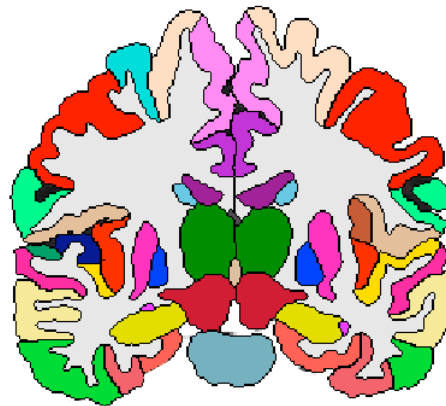


The Mindboggle project: feature-based brain labeling

arno klein

arno@binarybottle.com

asst. professor of clinical neurobiology
molecular imaging and neuropathology
columbia university

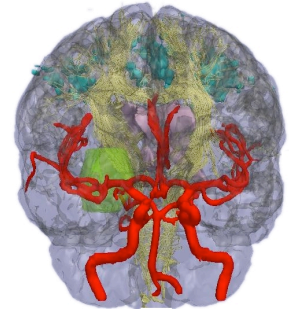


February 11, 2011
University of Pennsylvania

Why label brains?

Labels serve as a visual guide and teaching tool

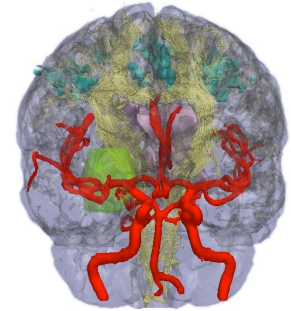
- teach brain anatomy
- guide neurosurgery



Why label brains?

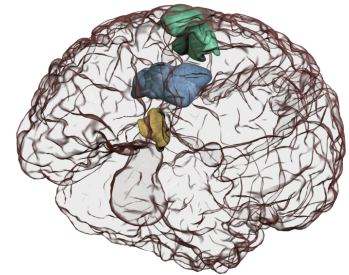
Labels serve as a visual guide and teaching tool

- teach brain anatomy
- guide neurosurgery



Labels break up data within a brain

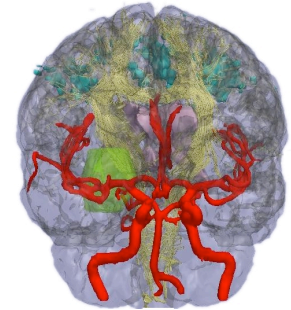
- assign results to brain regions
- quantify data by brain region



Why label brains?

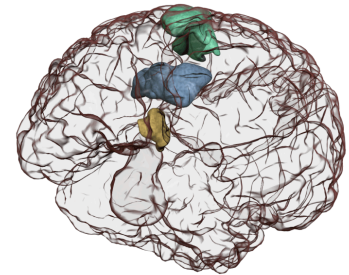
Labels serve as a visual guide and teaching tool

- teach brain anatomy
- guide neurosurgery



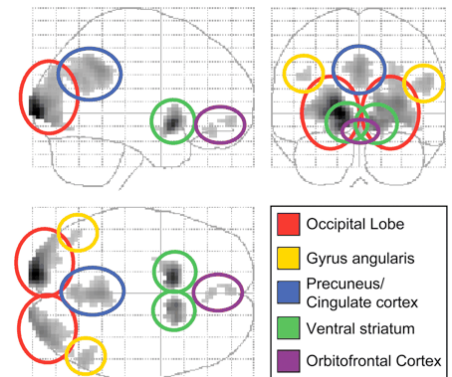
Labels break up data within a brain

- assign results to brain regions
- quantify data by brain region

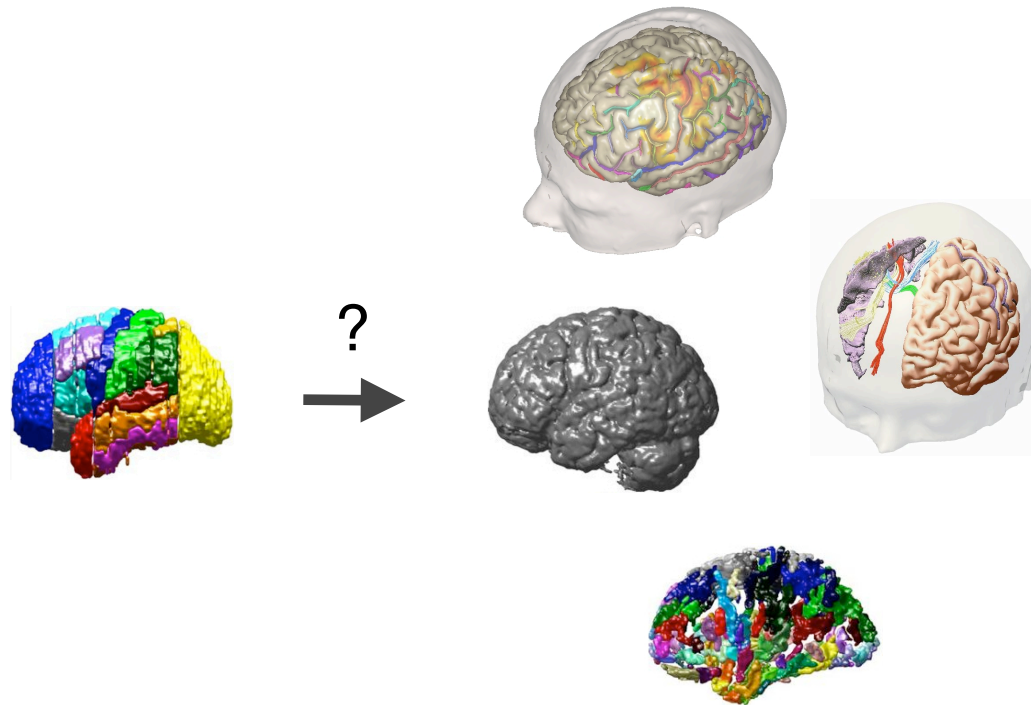


Labels establish correspondences across brains

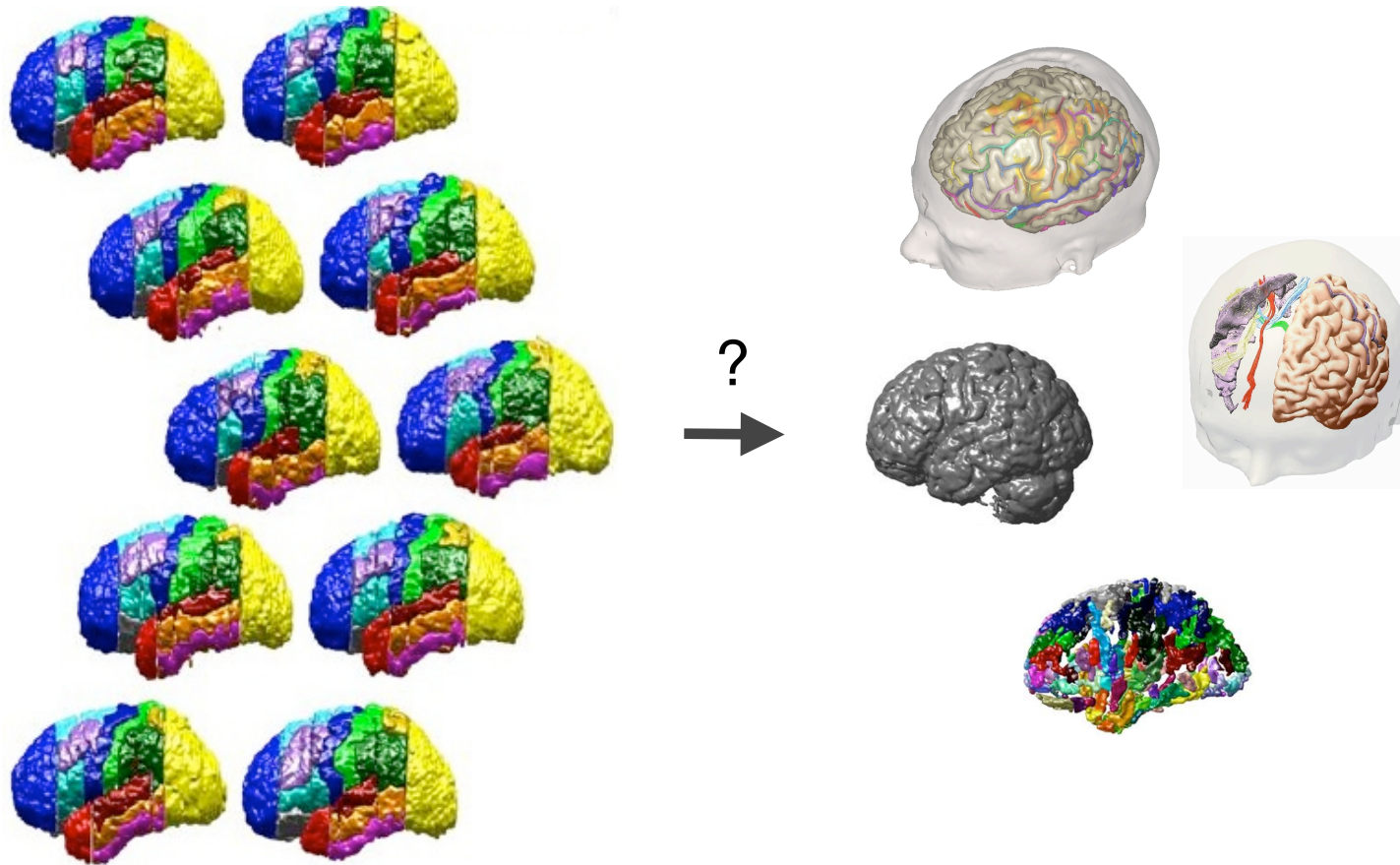
- compare individuals within and across studies
- communicate results with a common language



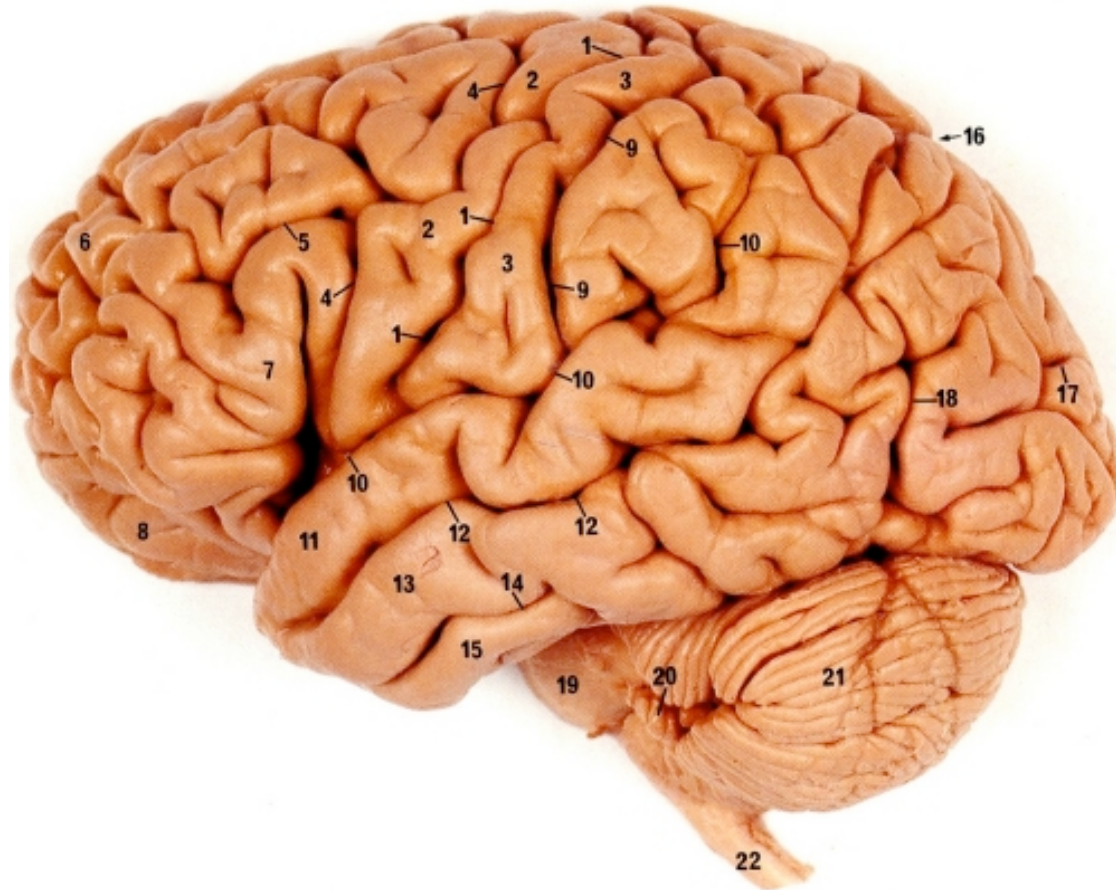
How should we label brains?

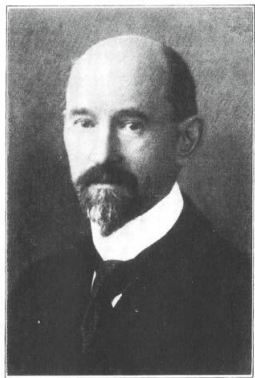


How should we label brains?



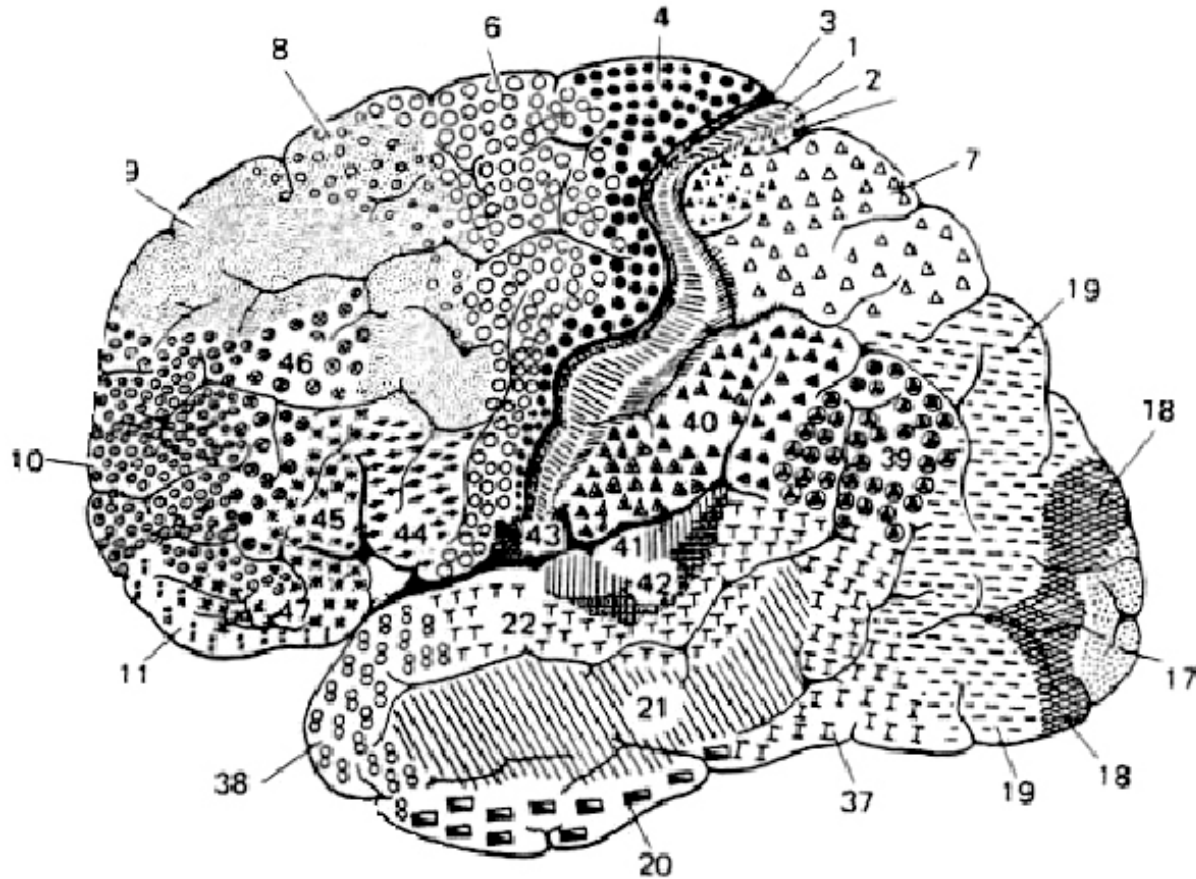
1. Manual labeling

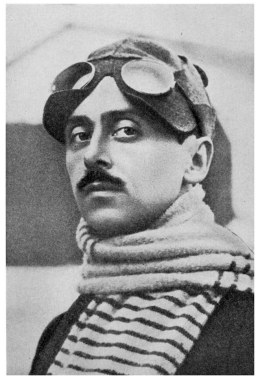




Korbinian Brodmann
(1868-1918)

Cytoarchitectonic boundaries





Cytoarchitectonic boundaries

Constantin von Economo
(1876-1931)

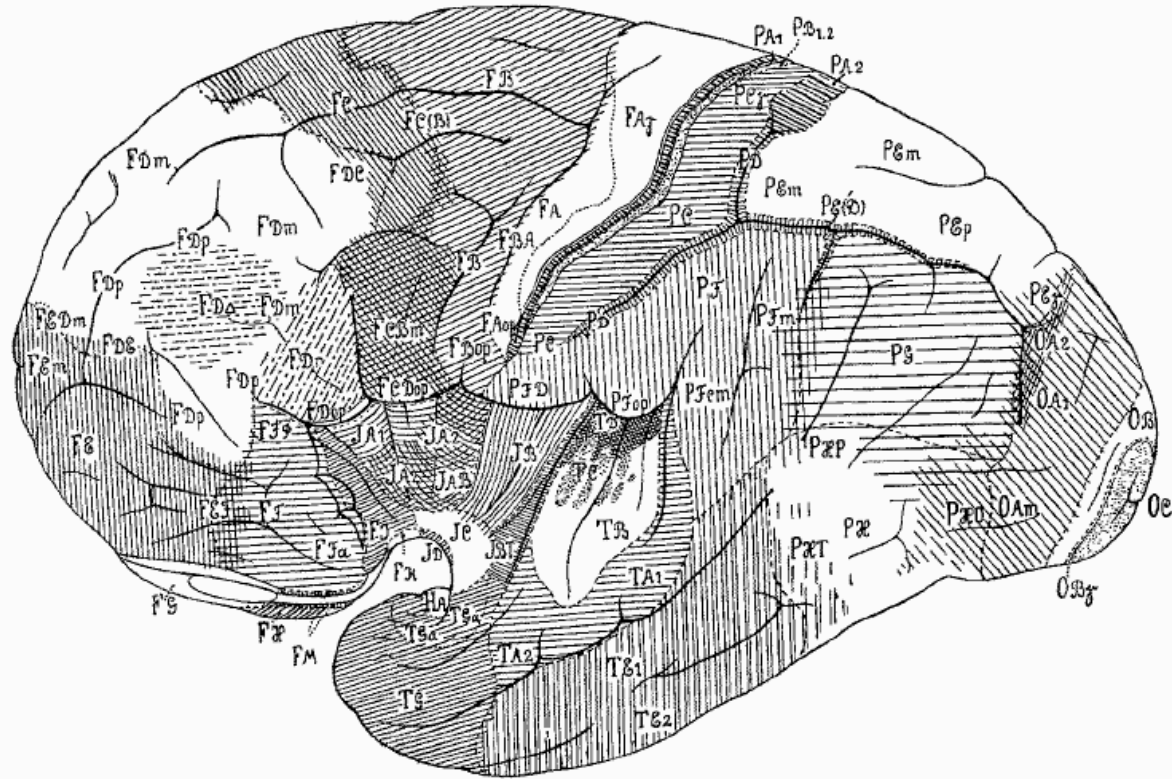
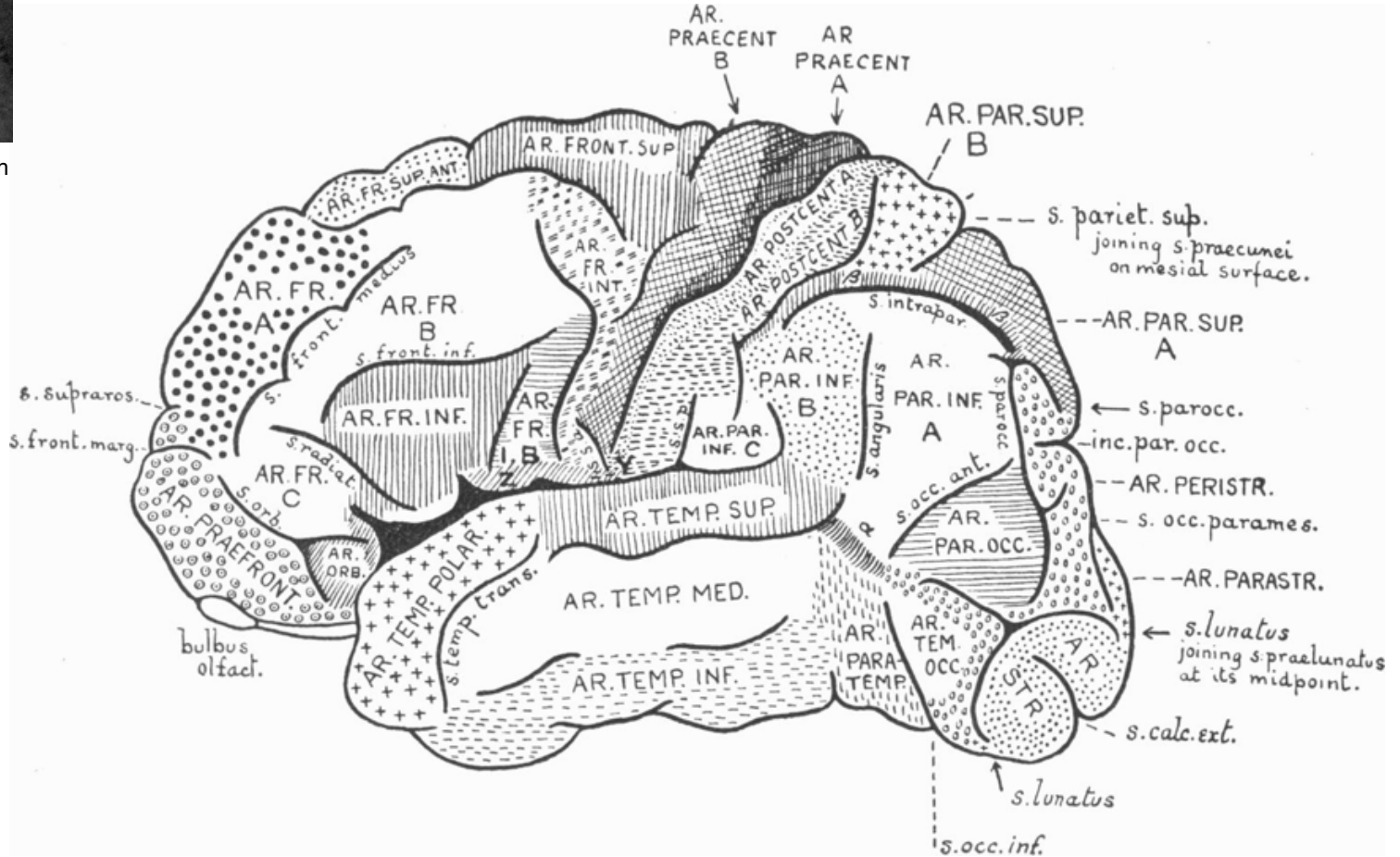


Abb. 3.

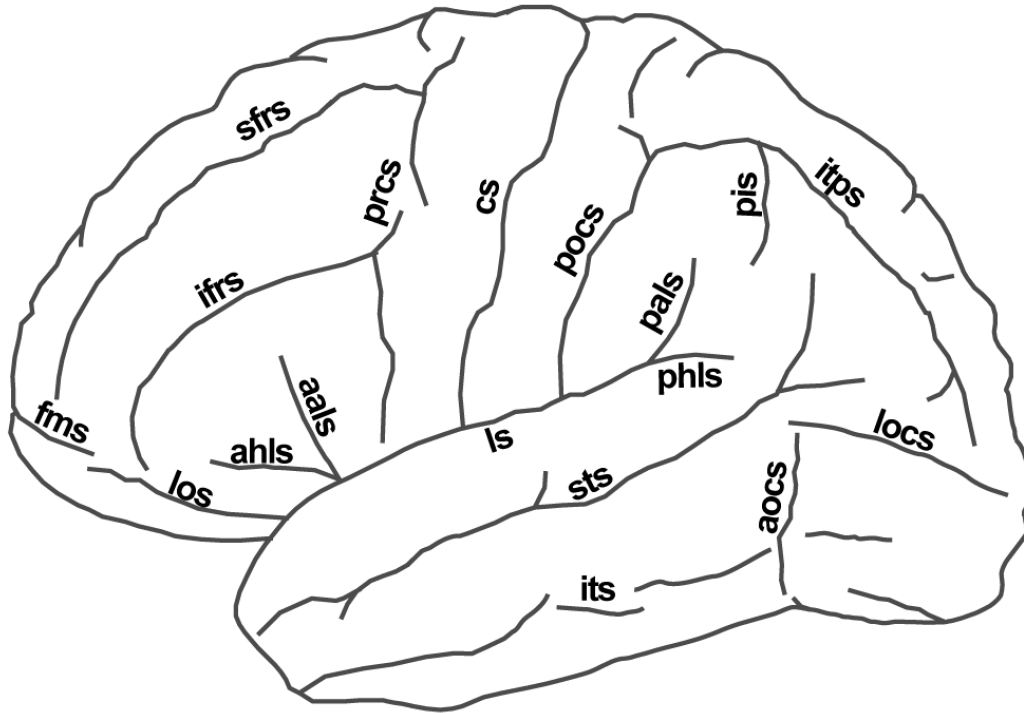


Sir Grafton Elliot Smith
(1871-1937)

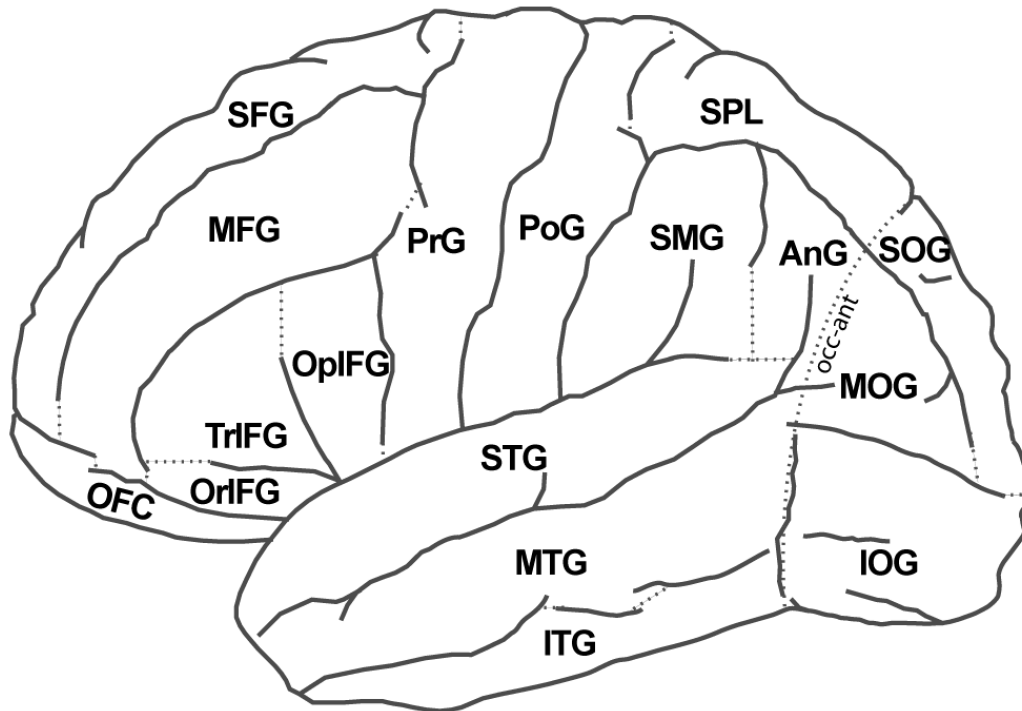
Topographic boundaries



Sulcus definitions

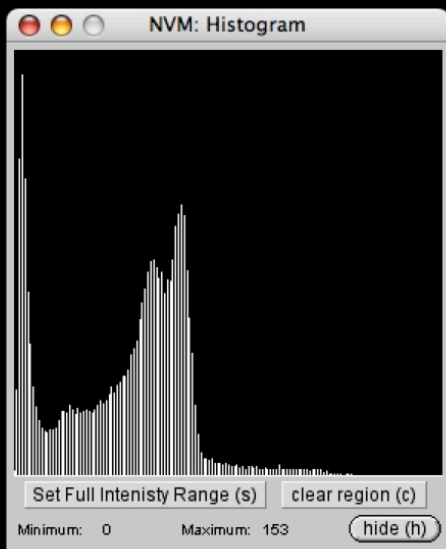


Gyrus definitions



File Edit Windows Tools Help

Segmenter Prefix: rjm **Right Hippocampus**



NVM: Landmarks
File Landmark Help

Choose a Scan:
10015_3

Choose a Landmark:

Right->Left X: 0
Superior-> Inferior Z: 0
Posterior-> Anterior Y: 0

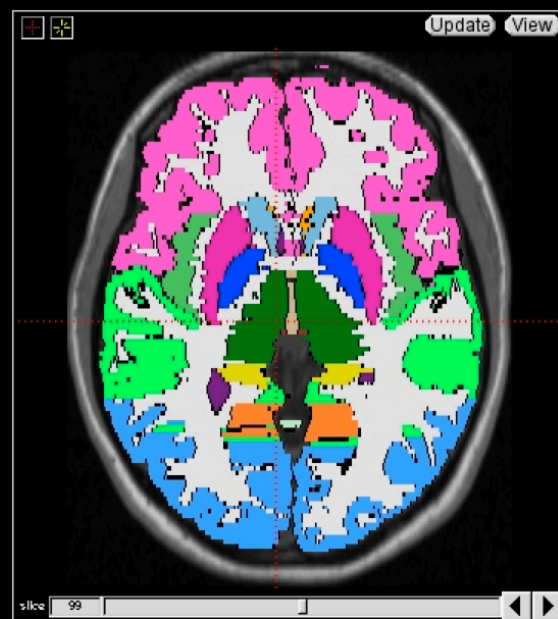
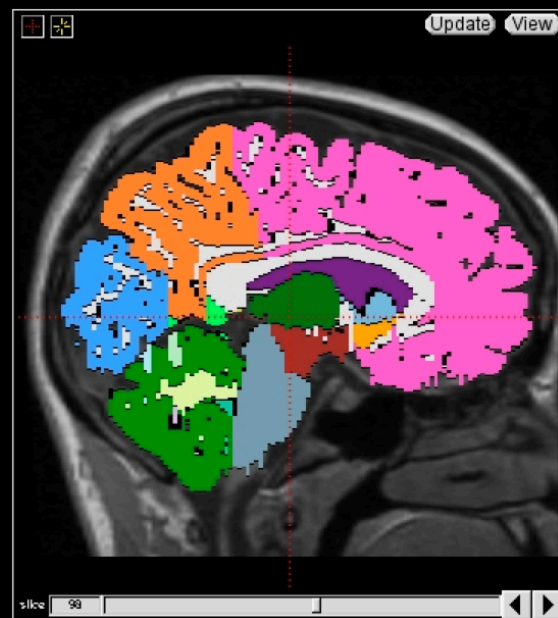
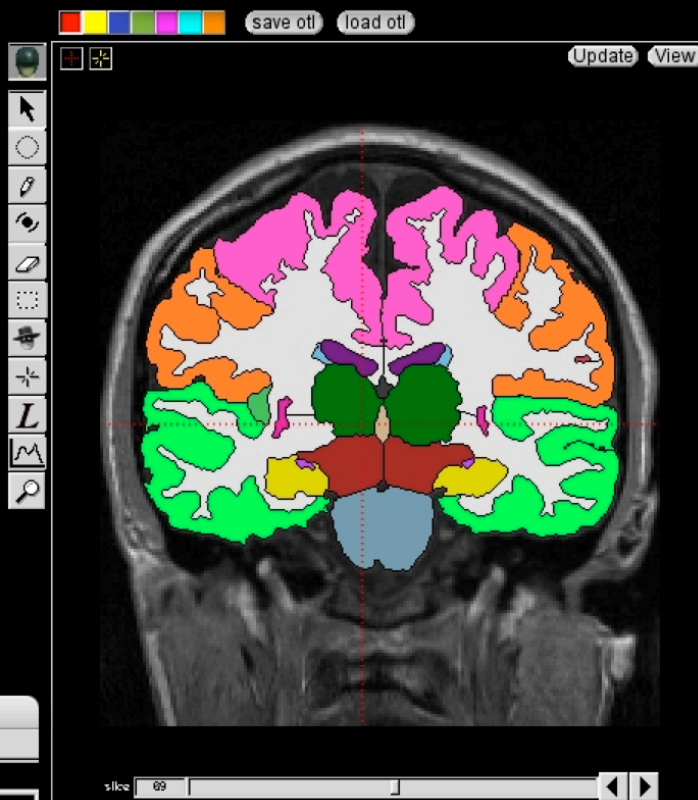
Review: Next Previous hide

NVM: Outline Labels
File Label Help

Assign current label when extracting

Choose Existing Label:
R-L Amygdala

Review: Next Previous hide

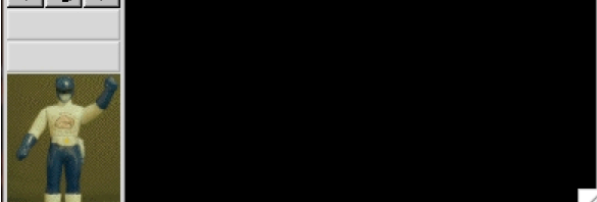


SegMentor v0.0
File Edit Actions Help

Ready to run: after last command (1 total) index.xml

Help
This SegMentor script will guide you through the segmentation

Prev. Next
To Do list
Hit the Enter key (with the main window selected and the mouse over an image) to begin...

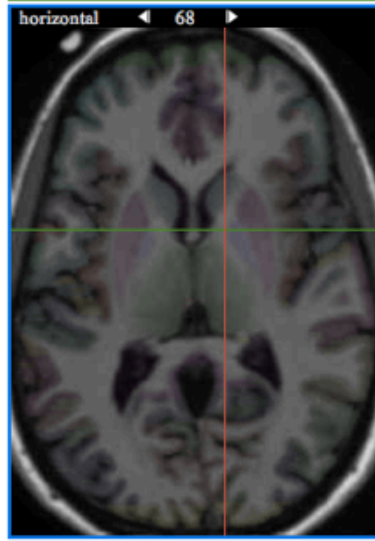
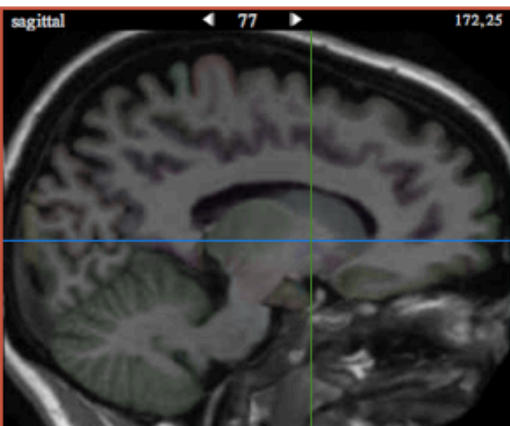
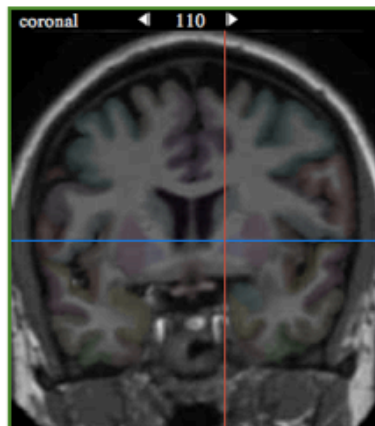


NVM: AutoContour slice 69
File AutoContour Help

Current	Contour	Label (and original intensity)
43		RoughBrain
13		Background-CSF
30		CSF-Gray
58		Gray-White



brainCOLOR Collaborative Open Labeling Online Resource



0% label opacity

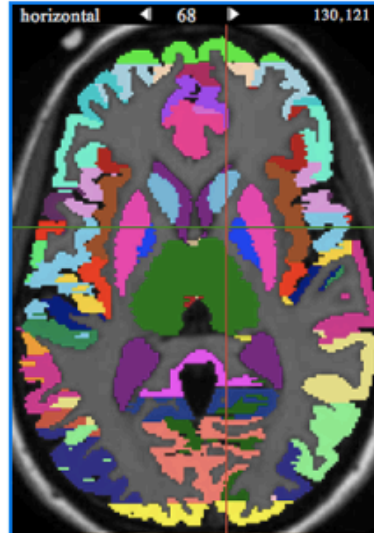
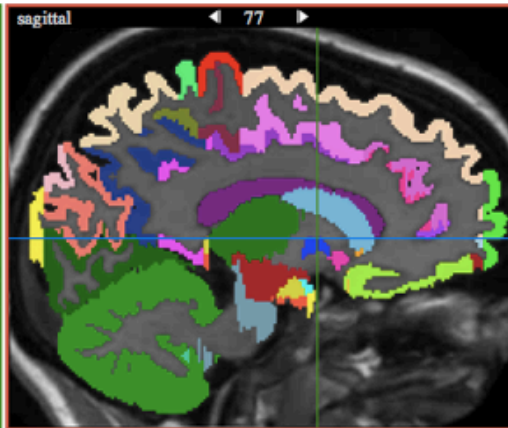
Coronal 110 labels (full label list):

- | | | |
|---|--|---------------|
| Left Lateral Ventricle | Right Lateral Ventricle | 3rd Ventricle |
| Left Caudate | Right Caudate | |
| Left Putamen | Right Putamen | |
| Left Pallidum | Right Pallidum | |
| Left Ventral DC | Right Ventral DC | |
| Left AIns anterior insula | Right Amygdala | |
| Left MCg middle cingulate gyrus | Right Insula | |
| Left PP planum polare | Right AIns anterior insula | |
| Left MFG middle frontal gyrus | Right PIns posterior insula | |
| Left PrG precentral gyrus | Right MCg middle cingulate gyrus | |
| Left PrG precentral gyrus | Right OpIFG opercular part of the inferior frontal gyrus | |
| Left Ent entorhinal area | Right MTG middle temporal gyrus | |
| Left ITG inferior temporal gyrus | Right ITG inferior temporal gyrus | |
| Left SFG superior frontal gyrus | Right FuG fusiform gyrus | |
| Left SMC supplementary motor cortex | Right SFG superior frontal gyrus | |
| Left OpIFG opercular part of the inferior frontal gyrus | Right MFG middle frontal gyrus | |
| Left CO central operculum | Right SMC supplementary motor cortex | |
| Left FuG fusiform gyrus | Right PrG precentral gyrus | |
| Left STG superior temporal gyrus | Right OpIFG opercular part of the inferior frontal gyrus | |
| Left MTG middle temporal gyrus | Right CO central operculum | |
| Left PIns posterior insula | Right PP planum polare | |
| | Right Ent entorhinal area | |
| | Right STG superior temporal gyrus | |



brainCOLOR

Collaborative Open Labeling Online Resource



100% label opacity

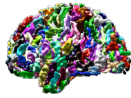


Coronal 110 labels (full label list):

- Left Lateral Ventricle
- Left Caudate
- Left Putamen
- Left Pallidum
- Left Ventral DC
- Left AIns anterior insula
- Left MCgG middle cingulate gyrus
- Left PP planum polare
- Left MFG middle frontal gyrus
- Left PrG precentral gyrus
- Left PrG precentral gyrus
- Left Ent entorhinal area
- Left ITG inferior temporal gyrus
- Left SFG superior frontal gyrus
- Left SMC supplementary motor cortex
- Left OpIFG opercular part of the inferior frontal gyrus
- Left CO central operculum
- Left FuG fusiform gyrus
- Left STG superior temporal gyrus
- Left MTG middle temporal gyrus
- Left Plns posterior insula

- Right Lateral Ventricle
- Right Caudate
- Right Putamen
- Right Pallidum
- Right Ventral DC
- Right Amygdala
- Right Insula
- Right AIns anterior insula
- Right Plns posterior insula
- Right MCgG middle cingulate gyrus
- Right OpIFG opercular part of the inferior frontal gyrus
- Right MTG middle temporal gyrus
- Right ITG inferior temporal gyrus
- Right FuG fusiform gyrus
- Right SFG superior frontal gyrus
- Right MFG middle frontal gyrus
- Right SMC supplementary motor cortex
- Right PrG precentral gyrus
- Right OpIFG opercular part of the inferior frontal gyrus
- Right CO central operculum
- Right PP planum polare
- Right Ent entorhinal area
- Right STG superior temporal gyrus

3rd Ventricle



An interactive tool for constructing optimal brain colormaps

Arno Klein NY State Psych. Inst. Columbia University, NY
 Andrew Worth Neuromorphometrics, Inc., MA
 Jason Tourville Cognitive & Neural Systems Boston University, MA
 Bennett Landman Vanderbilt University Nashville, TN
 Tito Dal Canton NY State Psych. Inst. Columbia University, NY
 Satrajit S. Ghosh Research Laboratory of Electronics, MIT, MA
 David Shattuck Laboratory of Neuromaging UCLA, CA

Goal

Create a software tool to aid in the construction of optimal colormaps for viewing and labeling brain anatomical images.

Background

The application of color is an indispensable means of visually distinguishing anatomical regions of the brain [1,2]. The need to facilitate visualization of brain images has become particularly urgent with the inception of large-scale anatomical labeling projects, such as the BrainCOLOR project, in which hundreds of MR images of the human brain are being manually labeled (www.braincolor.org/protocols) as well as viewed online (www.braincolor.org/roygbiv).

Choosing a color space

Our colormap must consist of colors that are perceptually distinct, so it is reasonable to choose a color space that is perceptually uniform, where a separation between two colors in the color space is as discriminable as the same separation between two other colors in the color space. The International Commission on Illumination (CIE) devised a color space in 1931 that facilitates color description (Fig.1). Subsequent improvements led to the CIELAB and CIELUV color spaces, as well as a cylindrical variant, CIELCh, all of which attempt to be perceptually uniform.

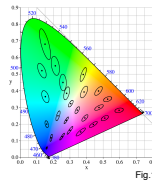


Fig.1

Constructing a colormap

We uniformly sampled hues from the CIELCh cylinder (Fig.2) at different distances from the center (70-100% chrominance) and at different lightness values. The resulting colormap (Fig.3) has the same number of colors as regions we wish to color (49 cortical regions in the BrainCOLOR protocol). We intend to expand the number of colors to include subcortical and other regions.

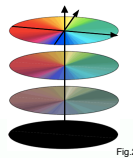


Fig.2

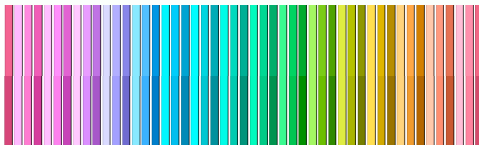
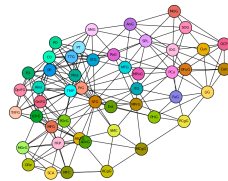


Fig.3

Maximizing discriminability of adjacent regions

To easily distinguish brain regions from each other, we wish to maximize the color difference between adjacent regions. We do this by constructing a graph that represents connections (edges) between regions (nodes).



We define an *optimal mapping* of colors as the combination of colors assigned to nodes with the maximum sum of color differences between connected nodes, defined by the CIE2000 Delta E measure (http://en.wikipedia.org/wiki/Color_difference). However, we cannot compute 8.8×10^{18} permutations of the 49 colors in the colormap, so we break up the brain into sublobes and compute the permutations of portions of the colormap at a time:

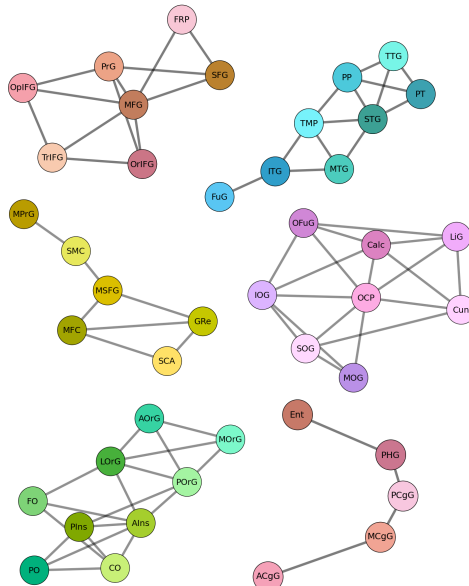
Frontal Lobe (7 A,B)
 - Lateral Surface (7)
 - Medial Surface (8)
 - Inferior Surface (4)
 Opercular Region (2)
 Insular Region (2)

Temporal Lobe (8)
 Lateral Surface (4)
 Supratemporal Surface (3)
 Inferior Surface (1)

Parietal lobe (6)
 Lateral Surface (4)
 Medial Surface (2)

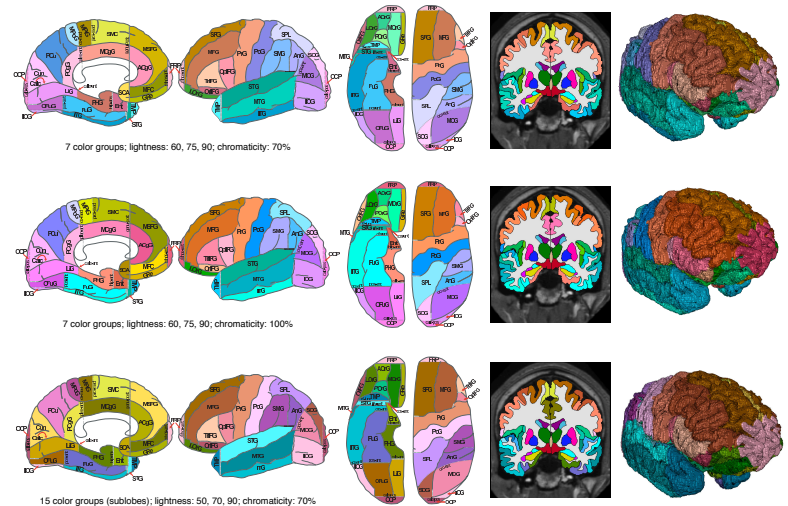
Occipital Lobe (8)
 Lateral Surface (4)
 Inferior Surface (1)
 Medial Surface (3)

Limbic Cortex (5)
 Cingulate Cortex (3)
 Medial Temporal Cortex (2)



Grouping regions by using similar colors

For labeling a brain image, maximum discriminability is paramount. For viewing a brain image, however, it is useful to visually group regions by assigning similar colors to the regions within a group. To create this visual grouping, we colored each subgraph with a neighborhood of colors (sampled within a wedge of the CIELCh cylinder).



Conclusion and Future Work

We have created Python software for creating and applying colormaps to brain images. Download the software from: <http://binarybottle.github.com/brainCOLORMap>. The software calls the Python libraries: *NumPy*, *NetworkX*, *Python-ColorMath*, *Matplotlib*, and *xird*, to:

1. convert a table of adjacent regions (an adjacency matrix) to a weighted graph of colored nodes
2. compute permutations of colors for each subgraph to optimally distinguish colors of adjacent regions
3. apply similar colors to predefined groups of regions

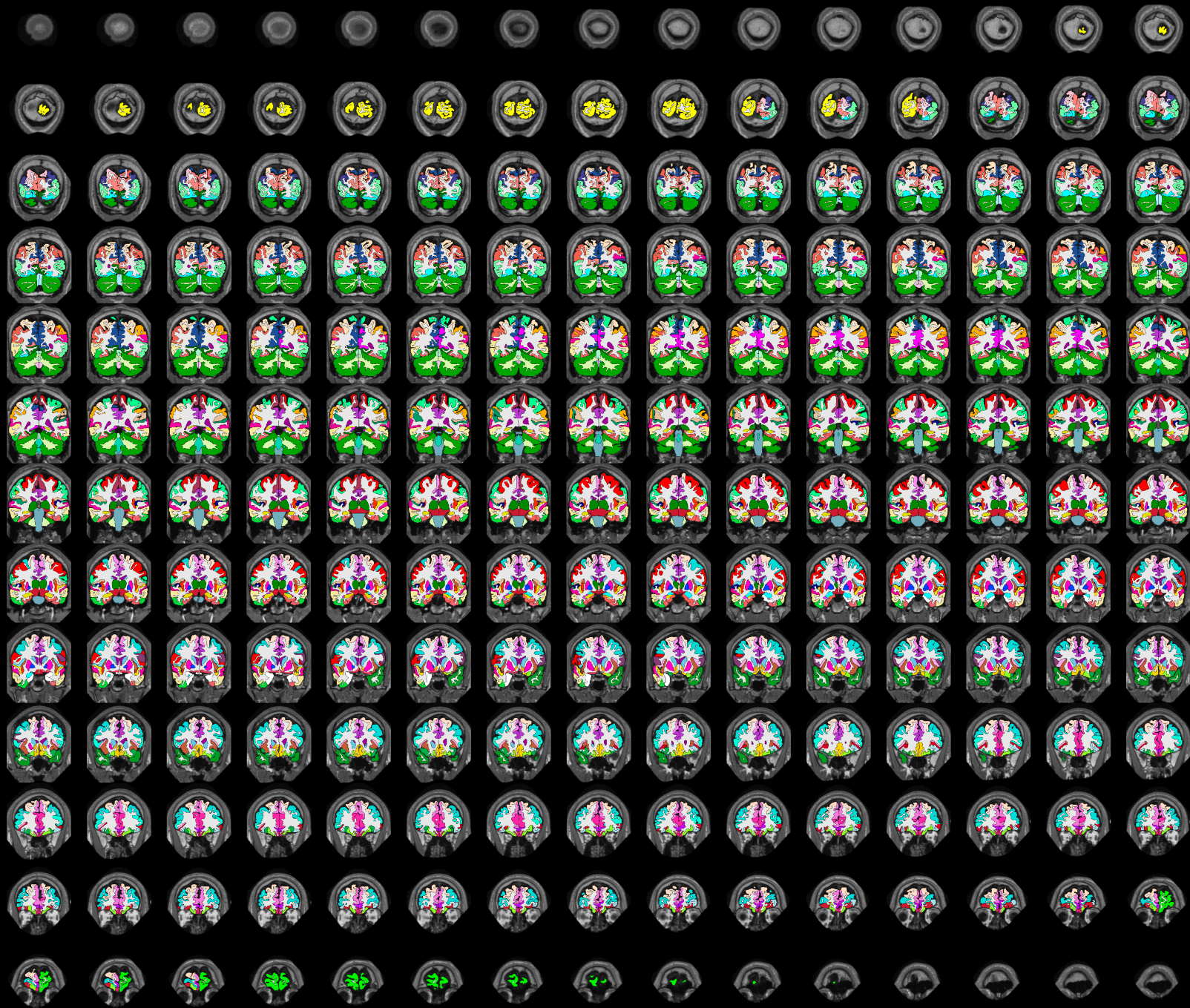
There are many improvements we could pursue. We have targeted the following for future work:

- ➔ Develop an optimization algorithm for whole graphs or larger subgraphs.
- ➔ Restrict the color space to colors that can be displayed; currently, the CIELCh colors that should appear different from one another may appear similar after conversion to sRGB colors.
- ➔ Seed color values to better control the range of colors for each group of regions.
- ➔ Accommodate visual concerns such as color blindness by adapting the colormap accordingly.

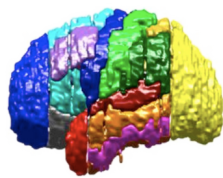


This project is funded as part of NIMH R01 MH084029-02 and NIMH R43 MH084358.

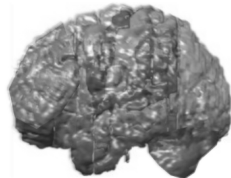
1. Schott GD (2010) Colored illustrations of the brain: some conceptual and contextual issues. *Neuroscience*. 16(5):808-16.
 2. Schyns P, Rodet C (2010) Portraits of the Mind: Visualizing the Brain from Antiquity to the 21st Century. *Alamy*. ISBN: 13: 978-0810960354
 CIE1931 figure: http://en.wikipedia.org/wiki/File:CIE1931_MacAdam.png
 CIELCh figure: <http://www.colorbasics.com/ColorSpace/>



2. Automated labeling



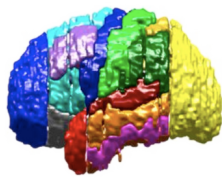
atlas



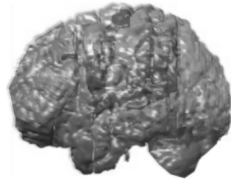
target



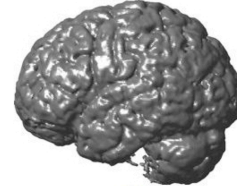
Convention: registration-based labeling



atlas

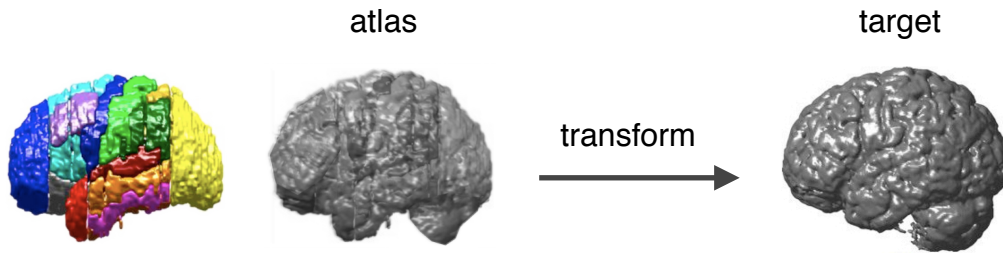


target



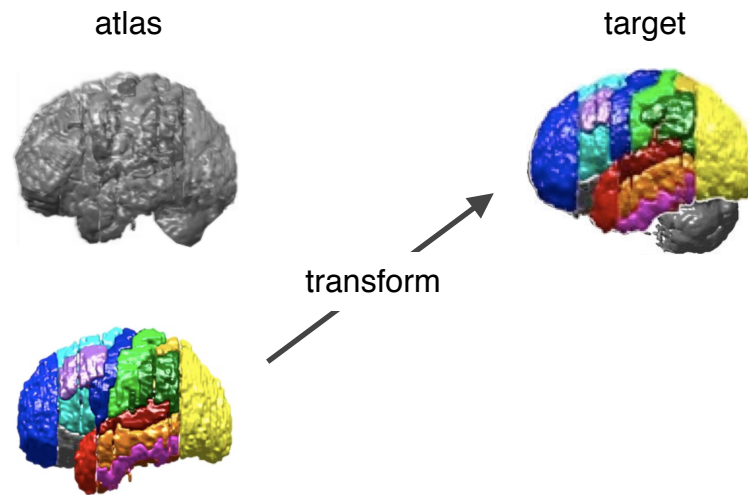
Convention: registration-based labeling

Step 1: compute the registration transform
from the atlas to the target

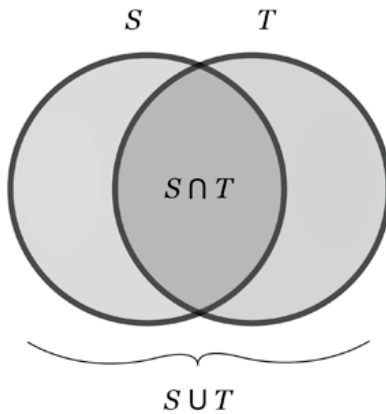


Convention: registration-based labeling

Step 2: apply the transform to the atlas labels



Evaluation



Target overlap: $TO = \frac{\sum_r |S_r \cap T_r|}{\sum_r |T_r|}$

Union overlap: $UO = \frac{\sum_r |S_r \cap T_r|}{\sum_r |S_r \cup T_r|}$

Mean overlap: $MO = 2 \frac{\sum_r |S_r \cap T_r|}{\sum_r (|S_r| + |T_r|)}$

Volume similarity: $VS = 2 \frac{\sum_r (|S_r| - |T_r|)}{\sum_r (|S_r| + |T_r|)}$

Hausdorff Distance Error:

$$DE(S, T) = \sum_r \text{mean}(\text{mean}_{s \in S_r}(\text{inf}_{t \in T_r}(d(s, t))), \text{mean}_{t \in T_r}(\text{inf}_{s \in S_r}(d(s, t))))$$

“Evaluation of 14 nonlinear deformation algorithms
applied to human brain MRI registration” NeuroImage (2009)

Algorithms: >14 software packages

Participants: 16 from 11 institutions

Data: 80 brains manually labeled according to 4 different
whole-brain labeling protocols (56 to 128 regions)

Registrations: Each algorithm applied >2,168 times (>45,000 total)

Evaluation: 8 measures (overlap, distance, volume similarity)

Analysis: 3 independent methods (ranking and statistical tests)

“Evaluation of 14 nonlinear deformation algorithms applied to human brain MRI registration”

NeuroImage (2009)

Software	Similarity metric	Transformation
SyN	CC	bi-directional diffeomorphism (D)
ART	nCC	FFD based on cubic splines (H, np)
IRTK	nMI	cubic B-splines
SPM5 DARTEL	multinomial model: congealing	FDM of viscosity field (Dc)
JRD-fluid	Jensen-Rényi divergence	viscous fluid; variational calculus (D)
Diffeomorphic Demons	SSD	displacement field (D, np)
FNIRT	SSD	cubic B-splines
ROMEO	displaced frame difference	local affine
ANIMAL	CC	local translations
SICLE	SSD	3-D Fourier series (D)
SPM5 Unified Segment	generative segmentation	discrete cosine transforms
SPM5 “SPM2-type”	MSD	discrete cosine transforms
SPM5 Normalize	MSD	discrete cosine transforms
AIR	MSD	5th-order polynomial warps
FLIRT (linear)	nCR	linear, rigid-body

n	= normalized	D	= diffeomorphic
CC	= cross-correlation	Dc	= diffeomorphic, constant over time
CR	= correlation ratio	FDM	= finite difference model
MI	= mutual information	FFD	= free-form deformation
MSD	= mean of squared differences	H	= homeomorphic
SSD	= sum of squared differences	np	= nonparametric

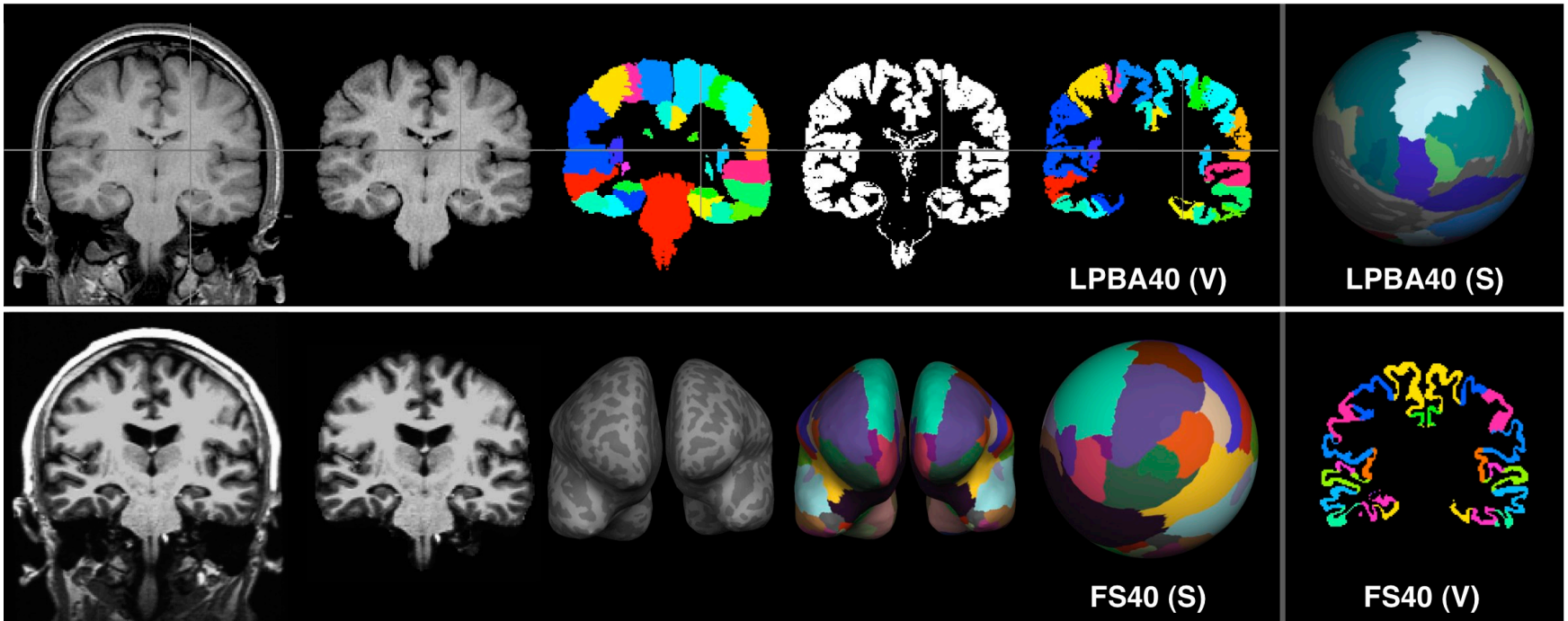
“Evaluation of volume-based and surface-based brain image registration methods”

NeuroImage (2010)

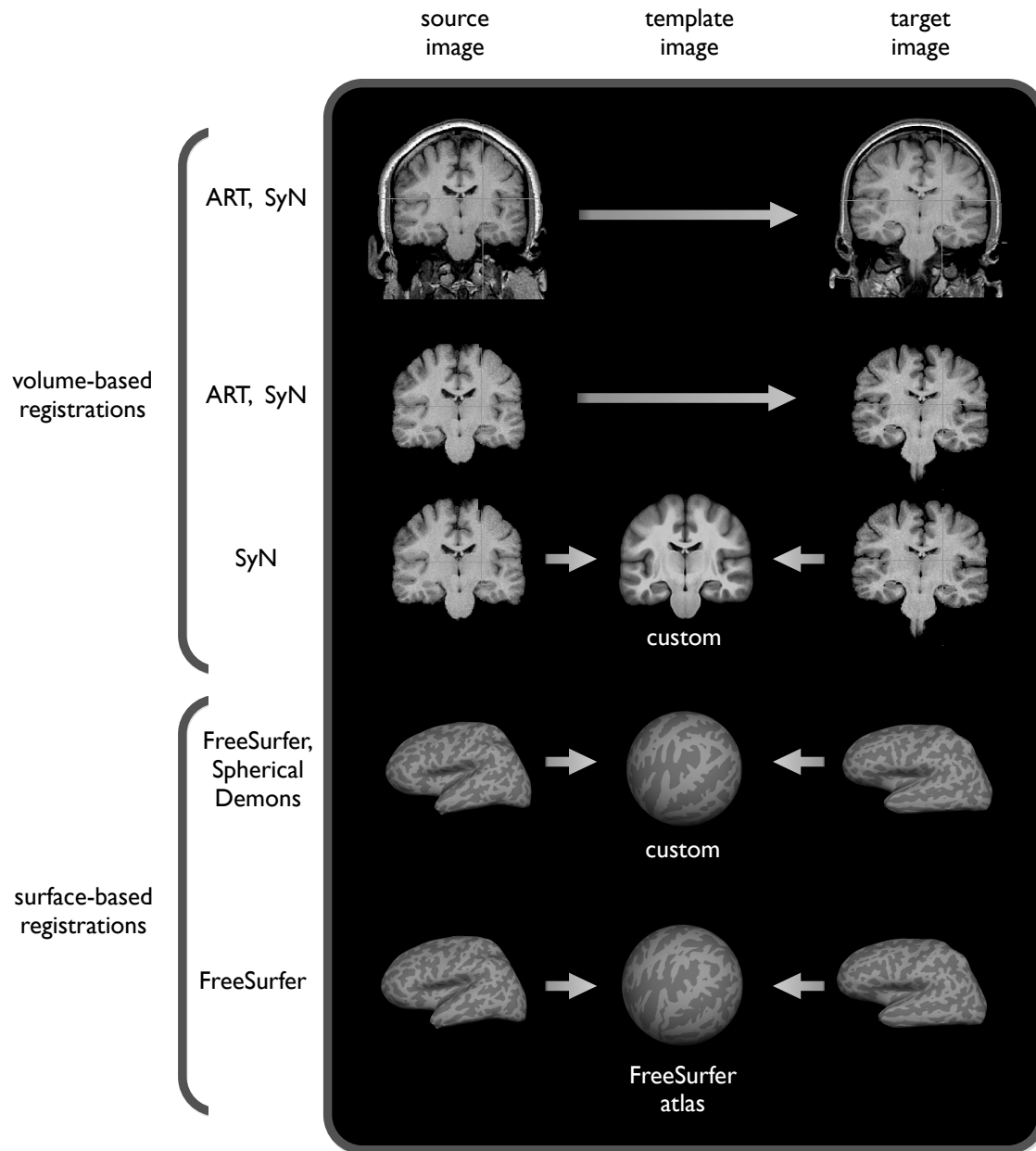
Collaborators:	Software:	Space:
Satrajit S. Ghosh		
Brian Avants, James C. Gee	ANTs (SyN)	volume
B.T.T. Yeo	Spherical Demons	surface
Bruce Fischl	FreeSurfer	surface
Babak Ardekani	ART	volume

- first study to compare volume and surface registration methods
- first study to compare whole-head and brain-only image registrations
- compares registration accuracy with and without custom templates
- >16,000 registrations
- 80 manually labeled brain images for evaluation
- 2 different brain labeling protocols

Volume and surface data
(labeled in volumes *and* surfaces)



registrations



Test 1: permutation test ranks

Volume registration methods

Rank 1 SyN with custom templates

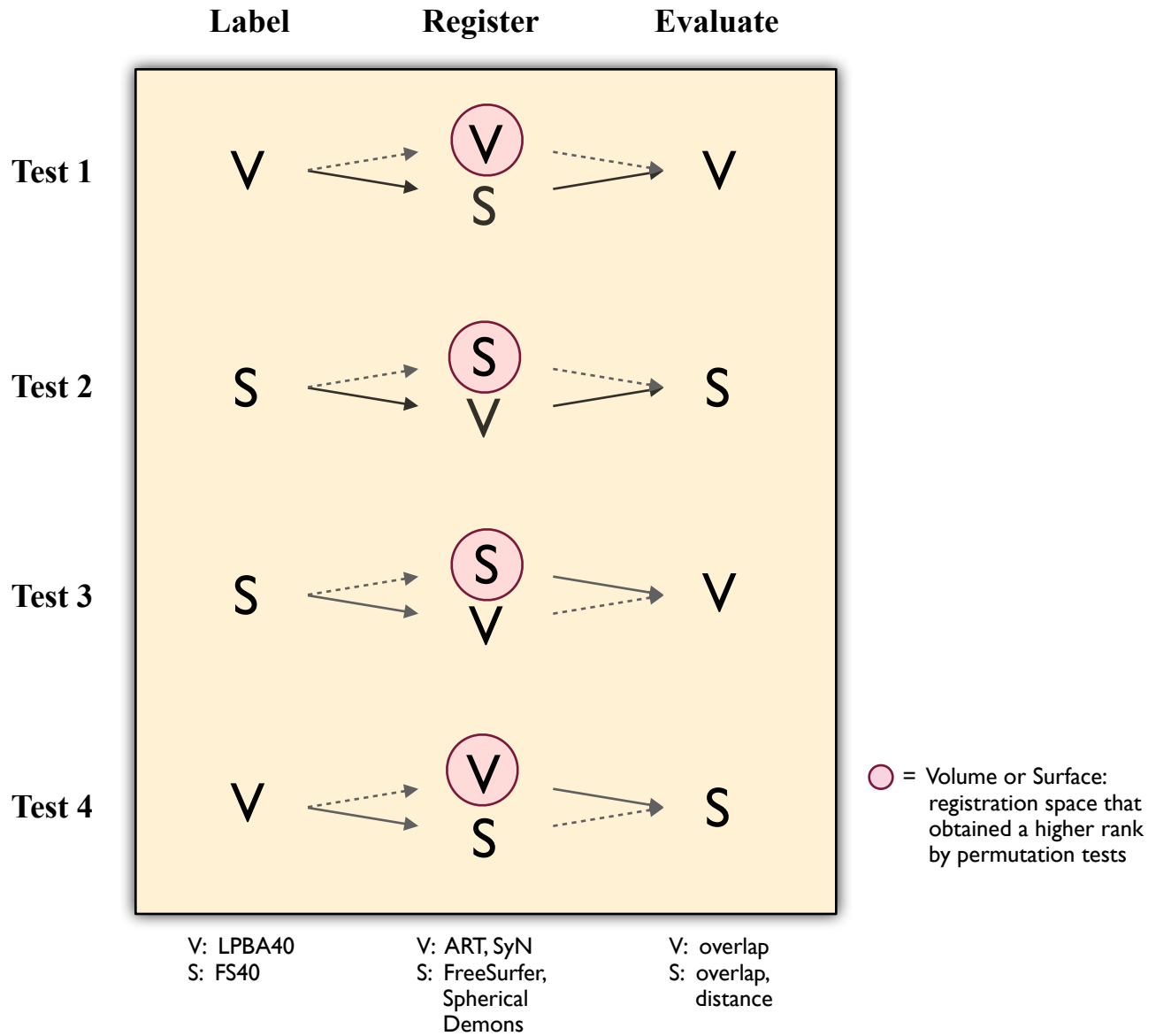
Rank 2 SyN
ART

Surface registration methods

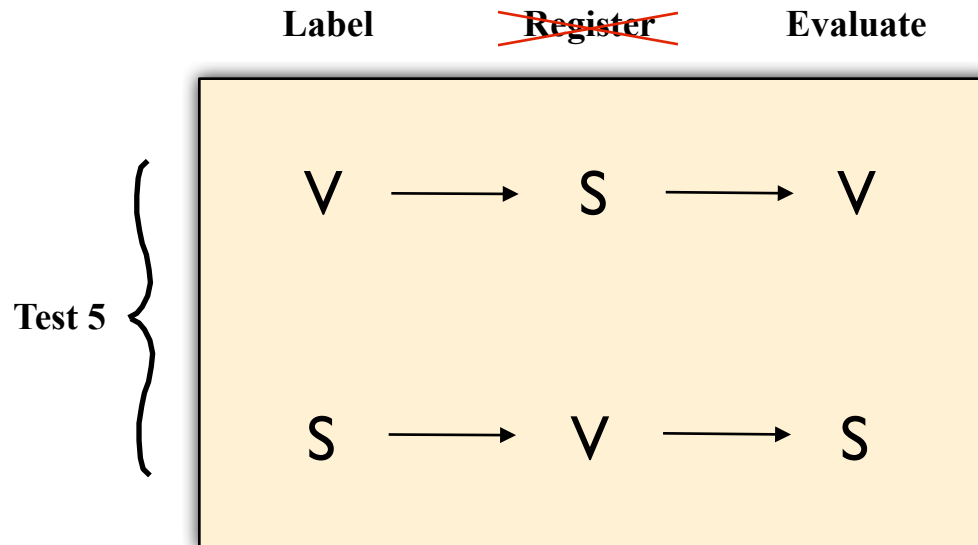
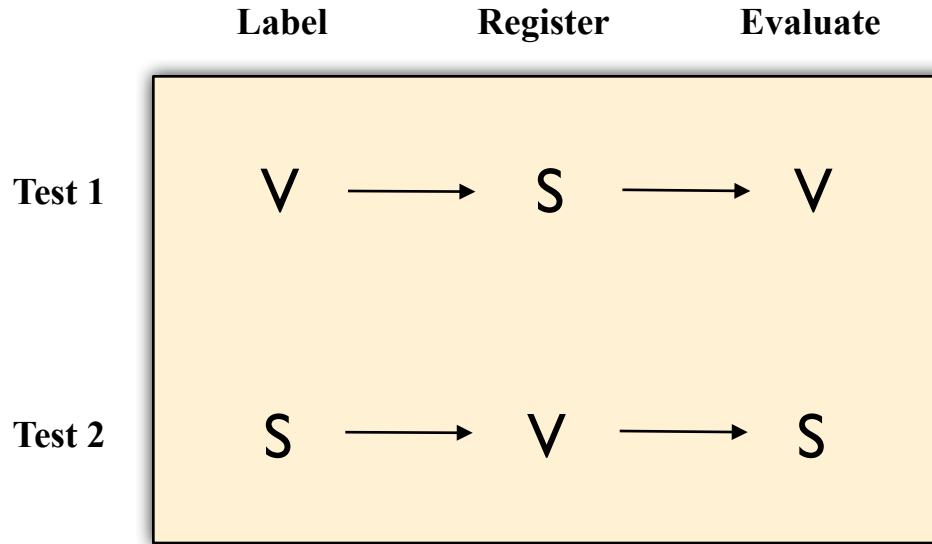
Rank 1 FreeSurfer with custom templates
Spherical Demons with custom templates

Rank 2 FreeSurfer with default atlas

Tests 1-4



Test 5: resampling



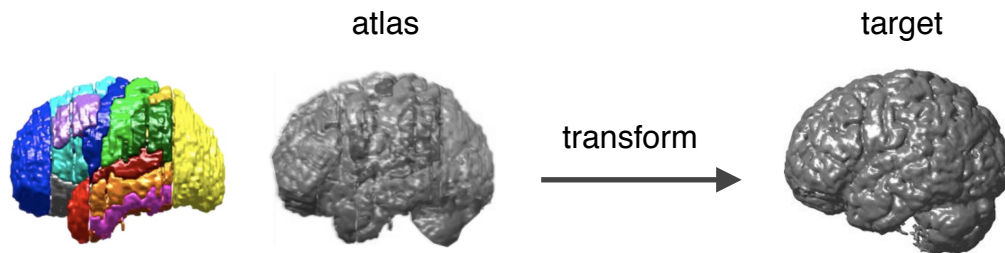
Test 5: resampling

1. Brain extraction aids volume registration.
2. Custom templates improve registration over direct pairwise registration.
3. Resampling volume labels on surfaces *or* surface labels on volumes precludes a fair comparison between surface and volume registration methods.

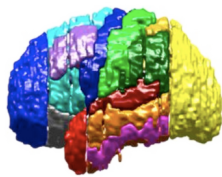
Recommendation:

Construct a custom template from a limited sample drawn from the same or a similar representative population, using the same algorithm used for registering brains to the template.

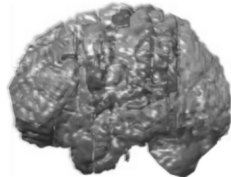
Convention: registration-based labeling



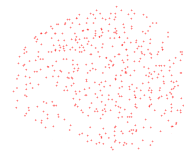
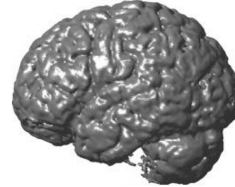
Mindboggle 2: feature-based labeling



atlas

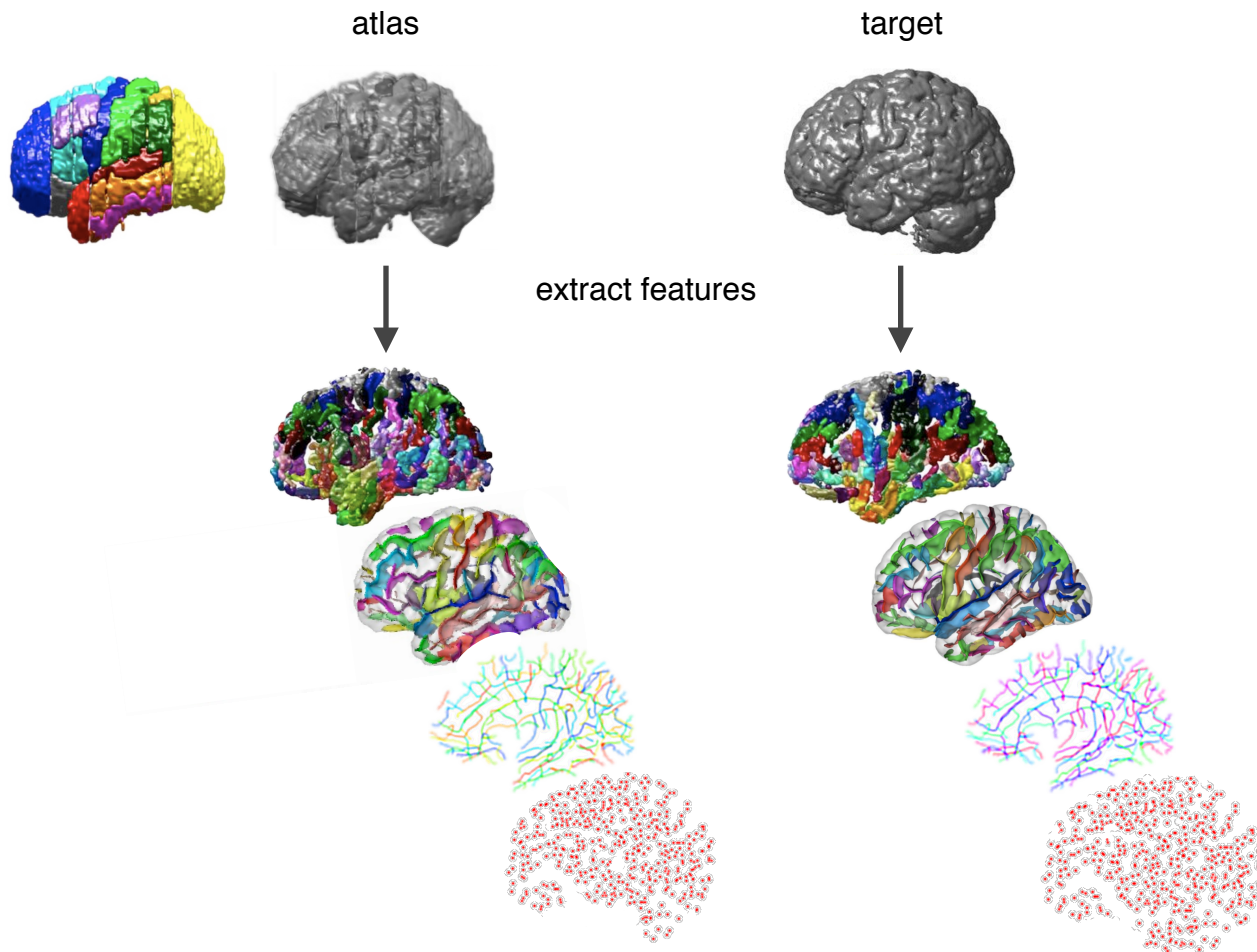


target



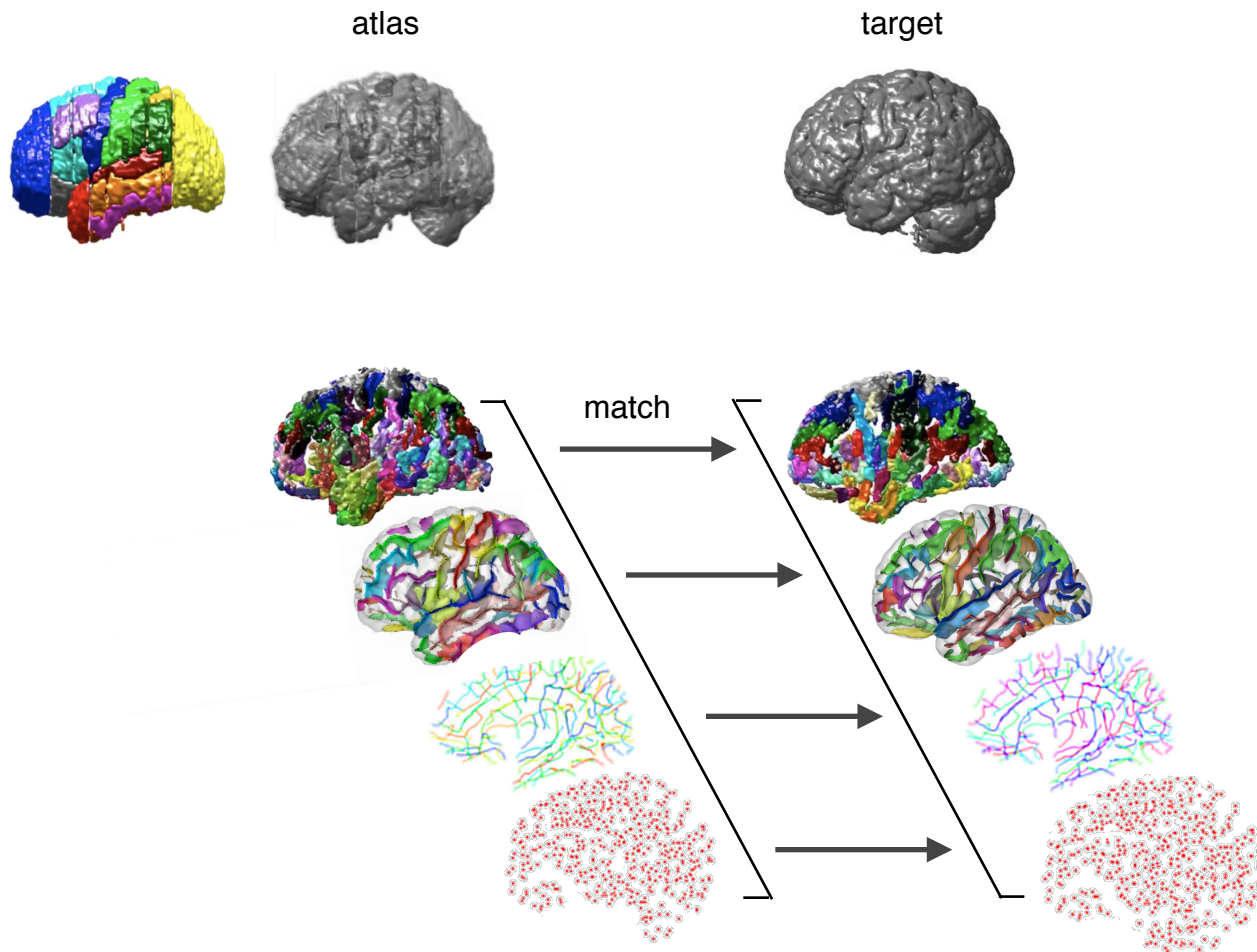
Mindboggle 2: feature-based labeling

Step 1: extract features



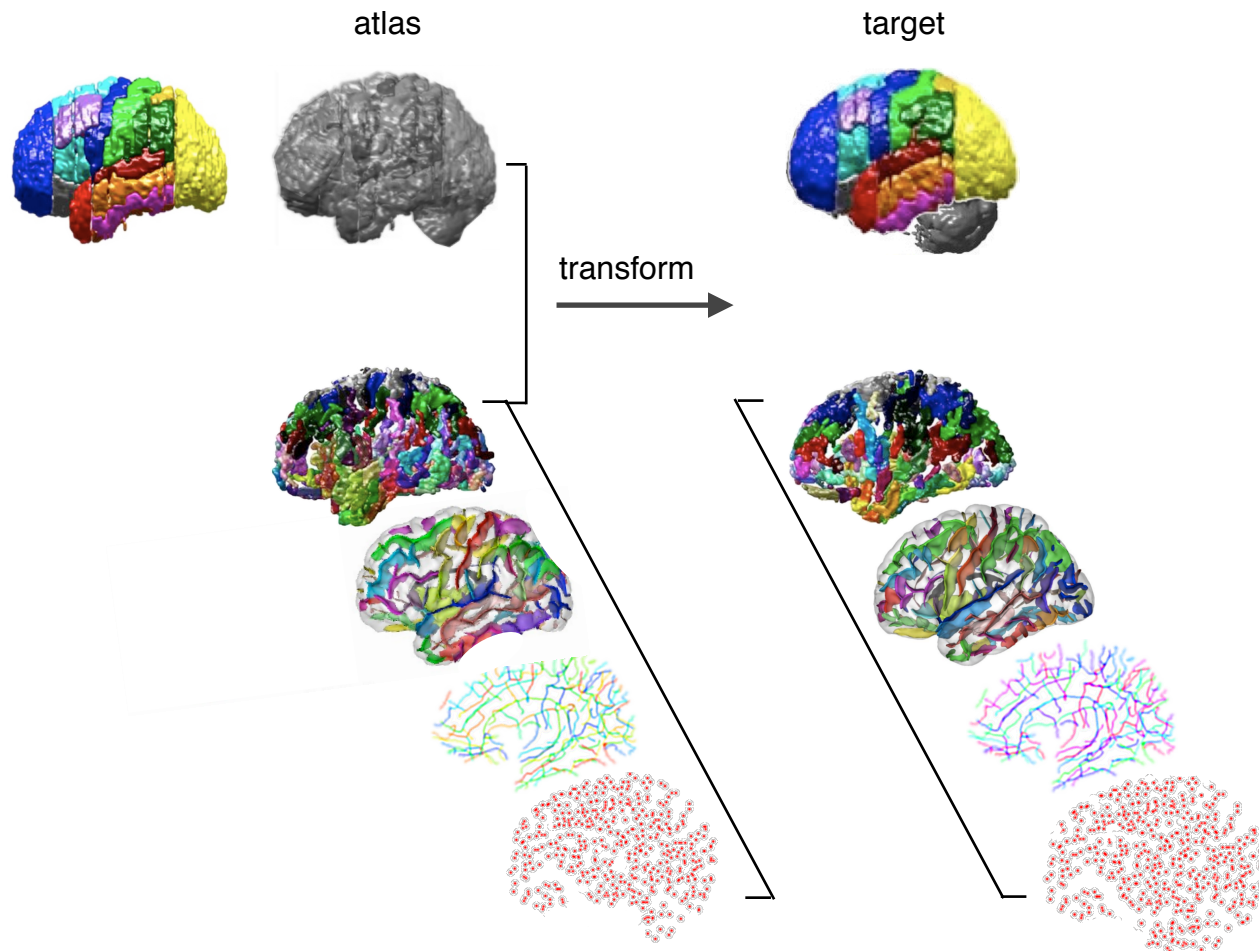
Mindboggle 2: feature-based labeling

Step 2: match atlas and target features



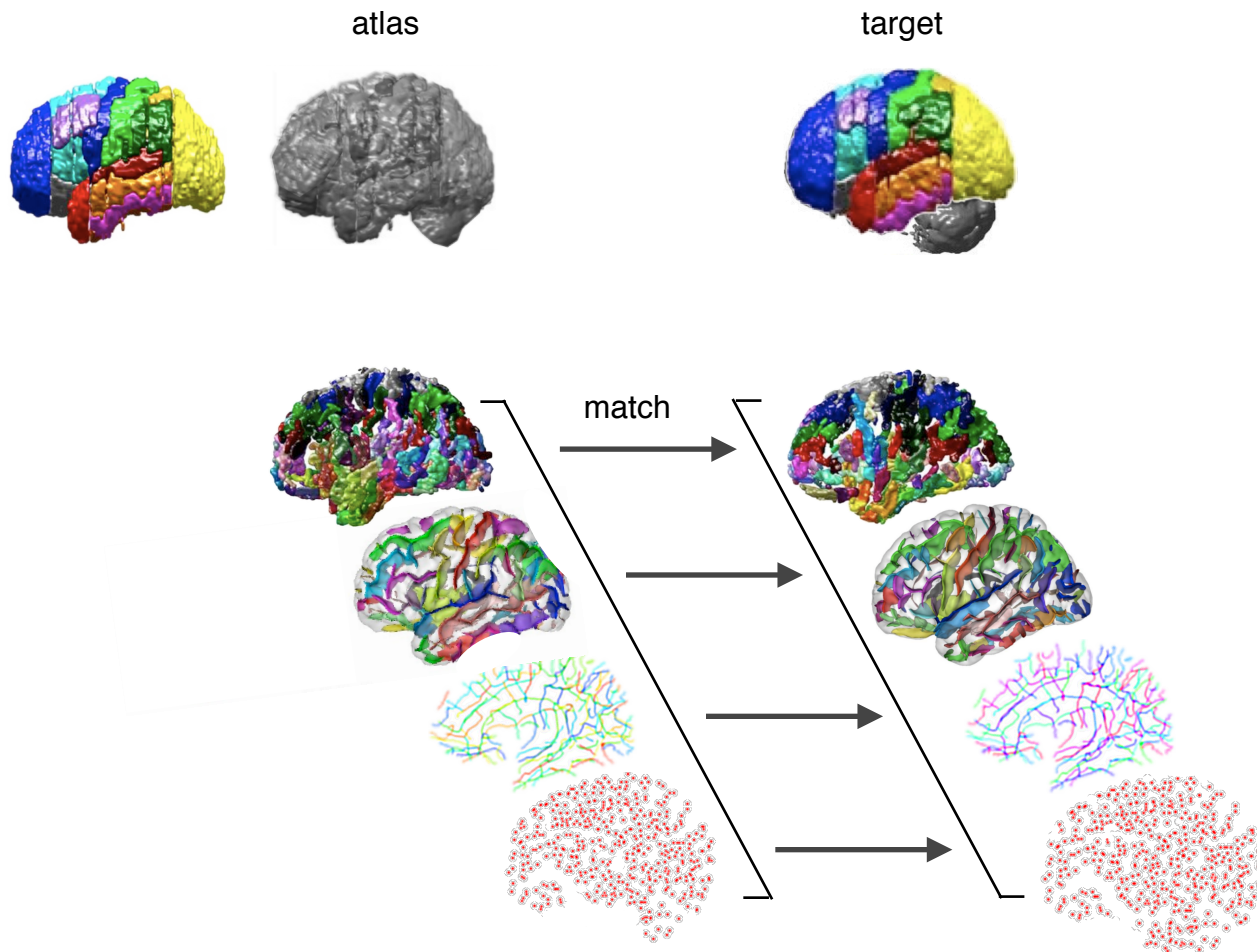
Mindboggle 2: feature-based labeling

Step 3: compute image + landmark-based registration transform from atlas to target



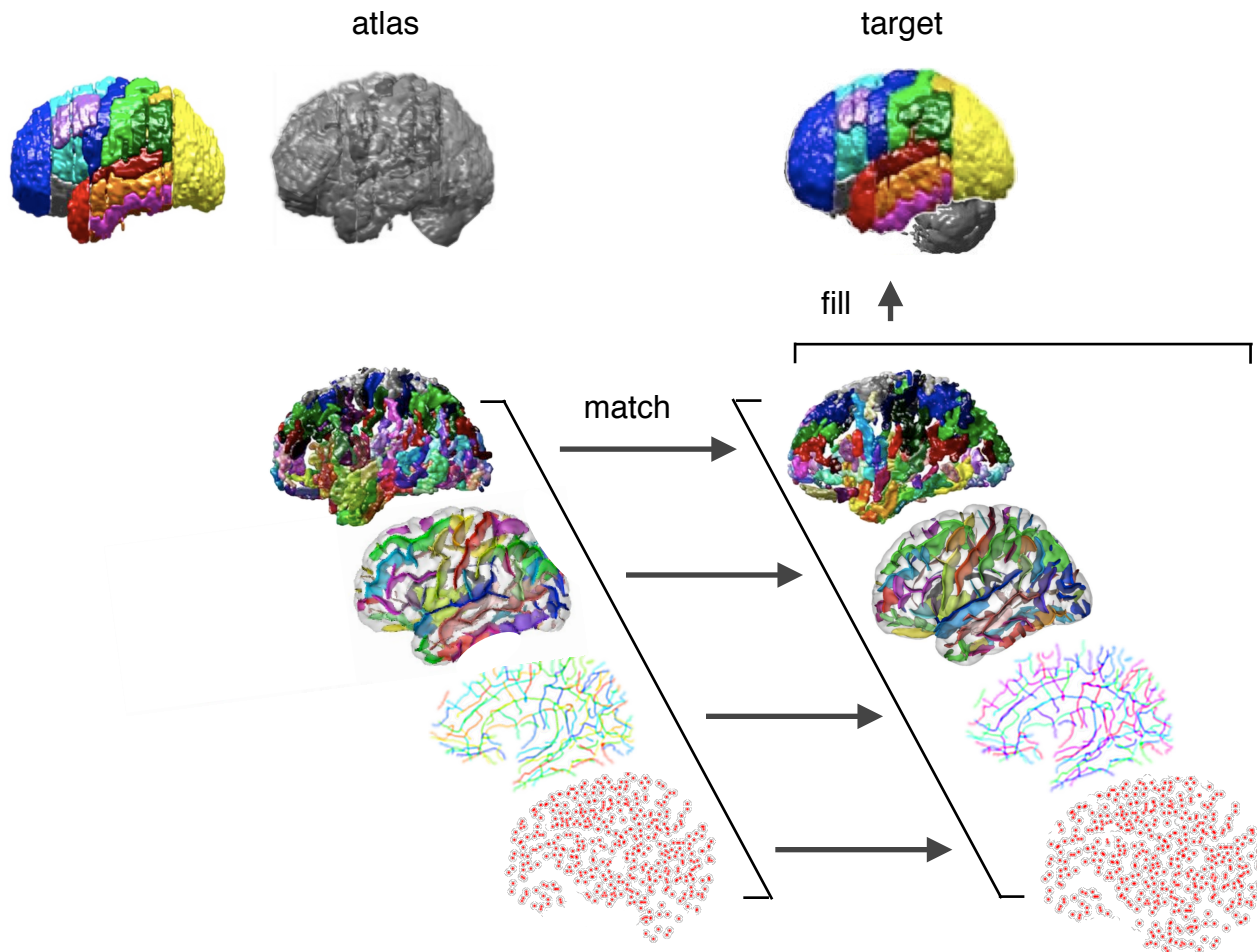
Mindboggle 2: feature-based labeling

Step 2: or match...

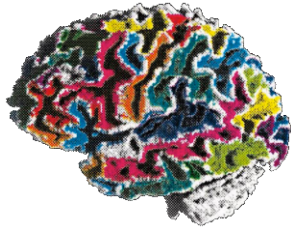


Mindboggle 2: feature-based labeling

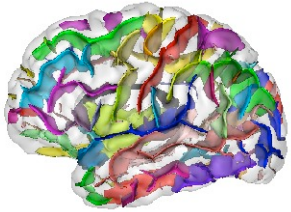
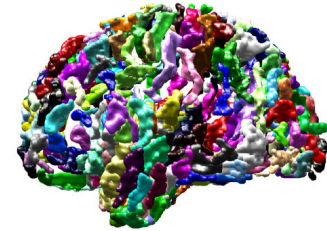
Step 3: then propagate labels within inferred label boundaries?



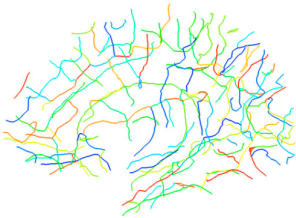
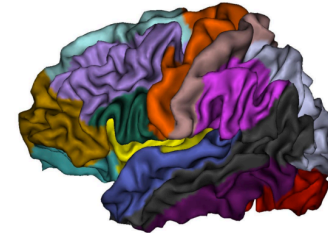
Candidate features



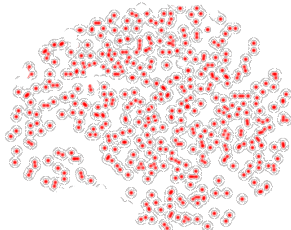
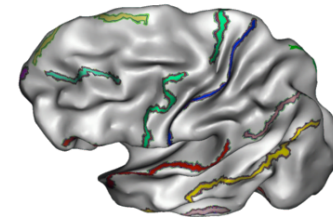
3-D:
labeled **regions** (manual)
sulcal **basins**
sulcal **skeletons**



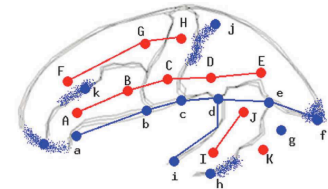
2-D:
sulcal **ribbons**
gyral **surfaces**



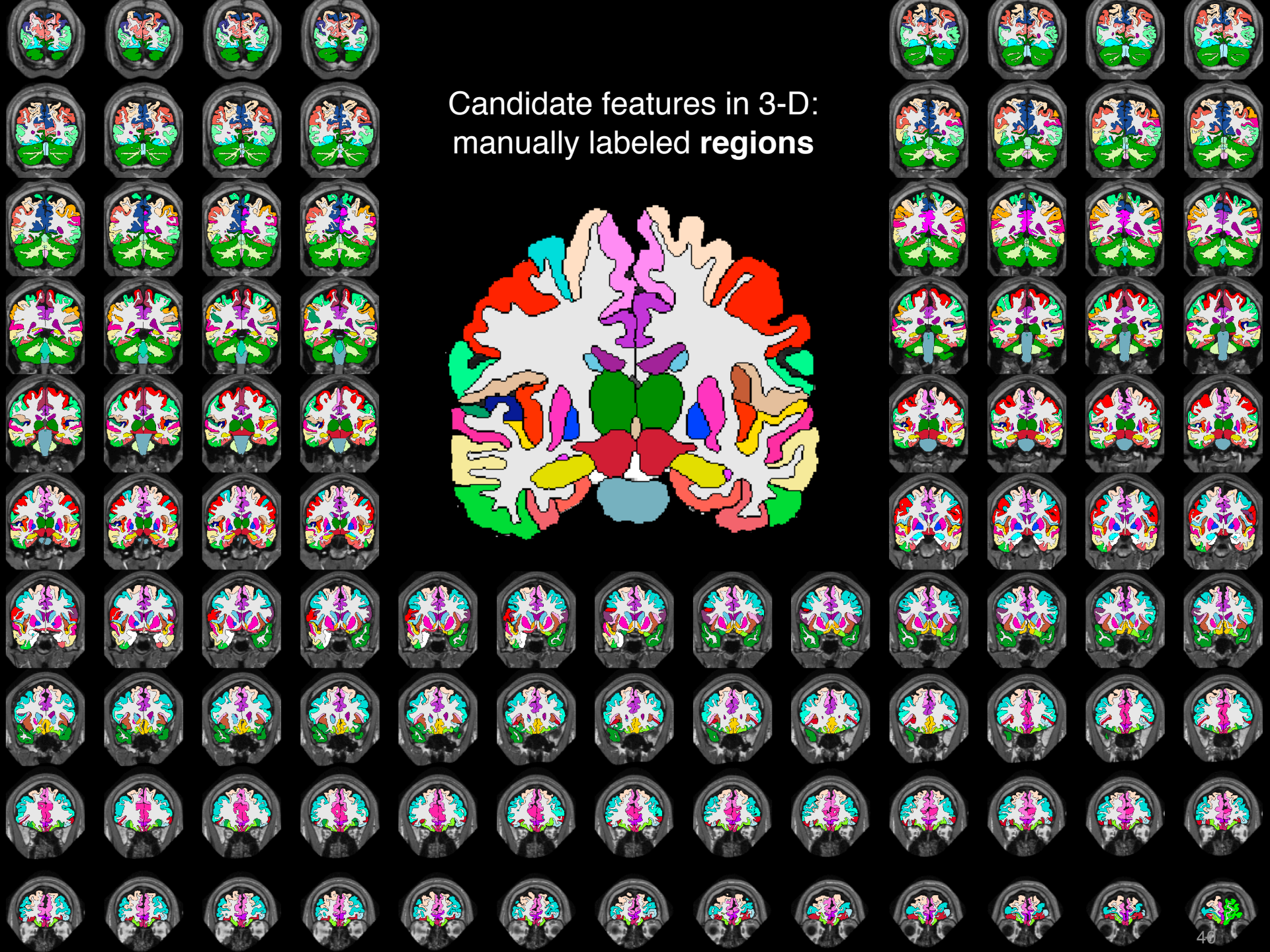
1-D:
sulcal
& gyral **curves**

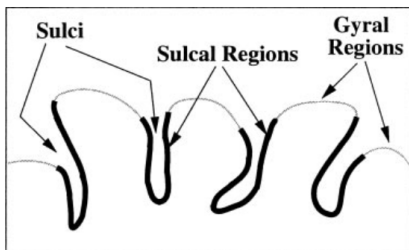


0-D:
SIFT **points**
sulcal **pits**

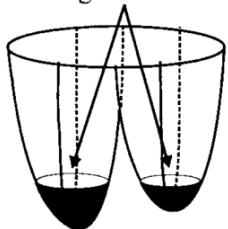


Candidate features in 3-D:
manually labeled regions

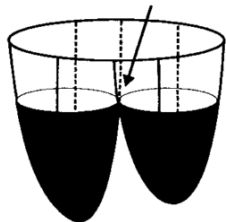




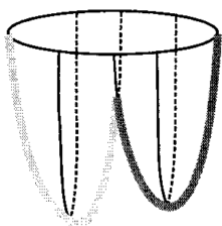
catchment basins begin filling with water



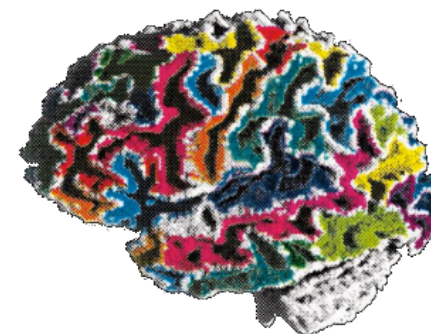
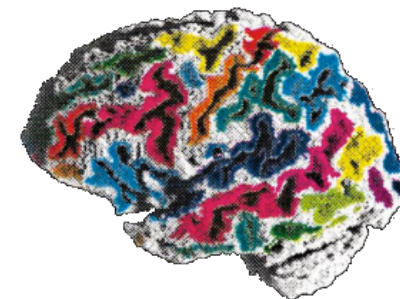
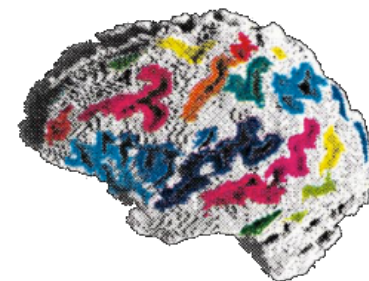
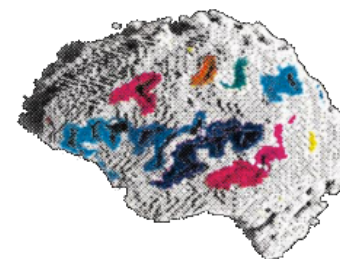
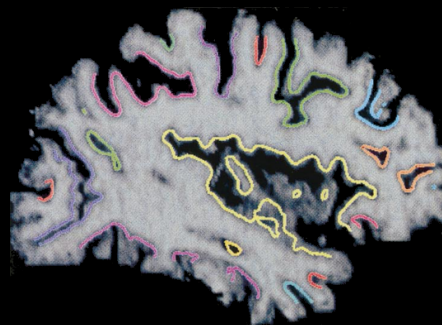
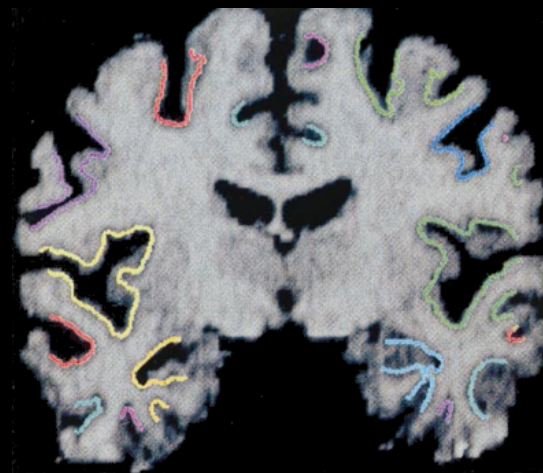
watershed line forms here



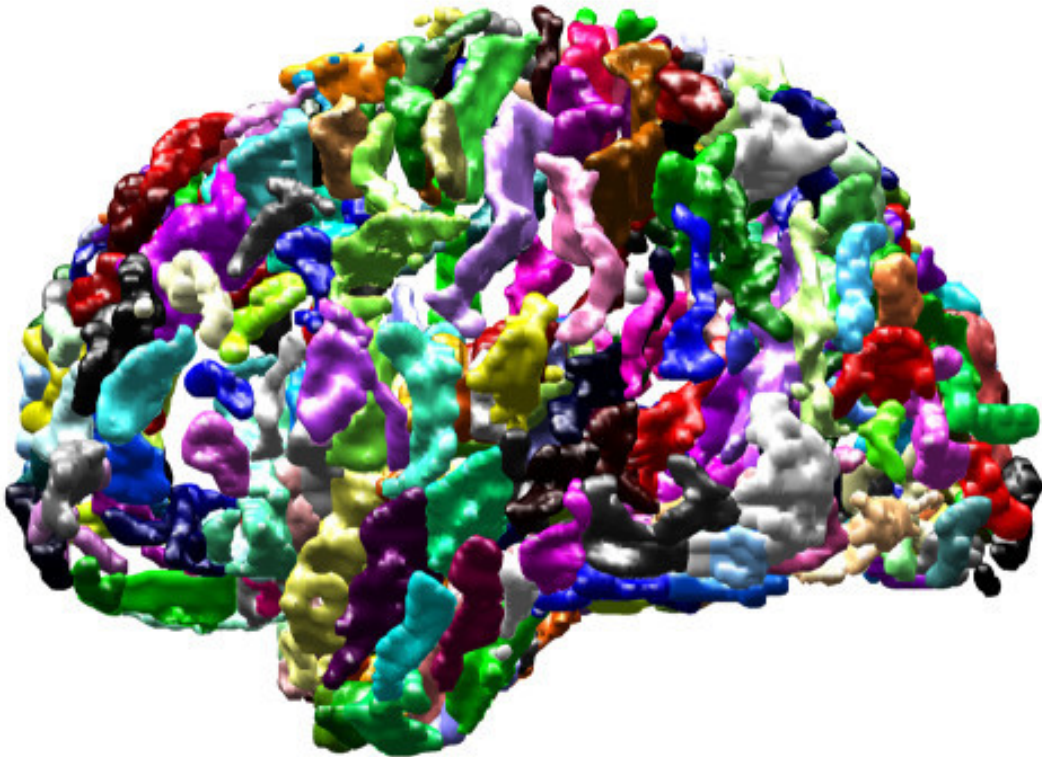
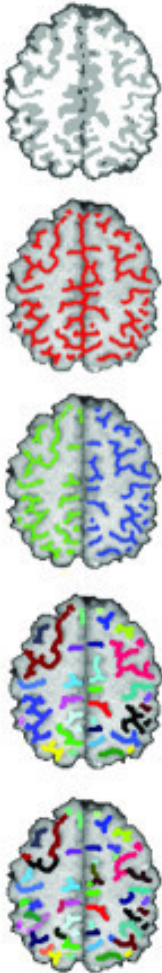
ideal segmentation



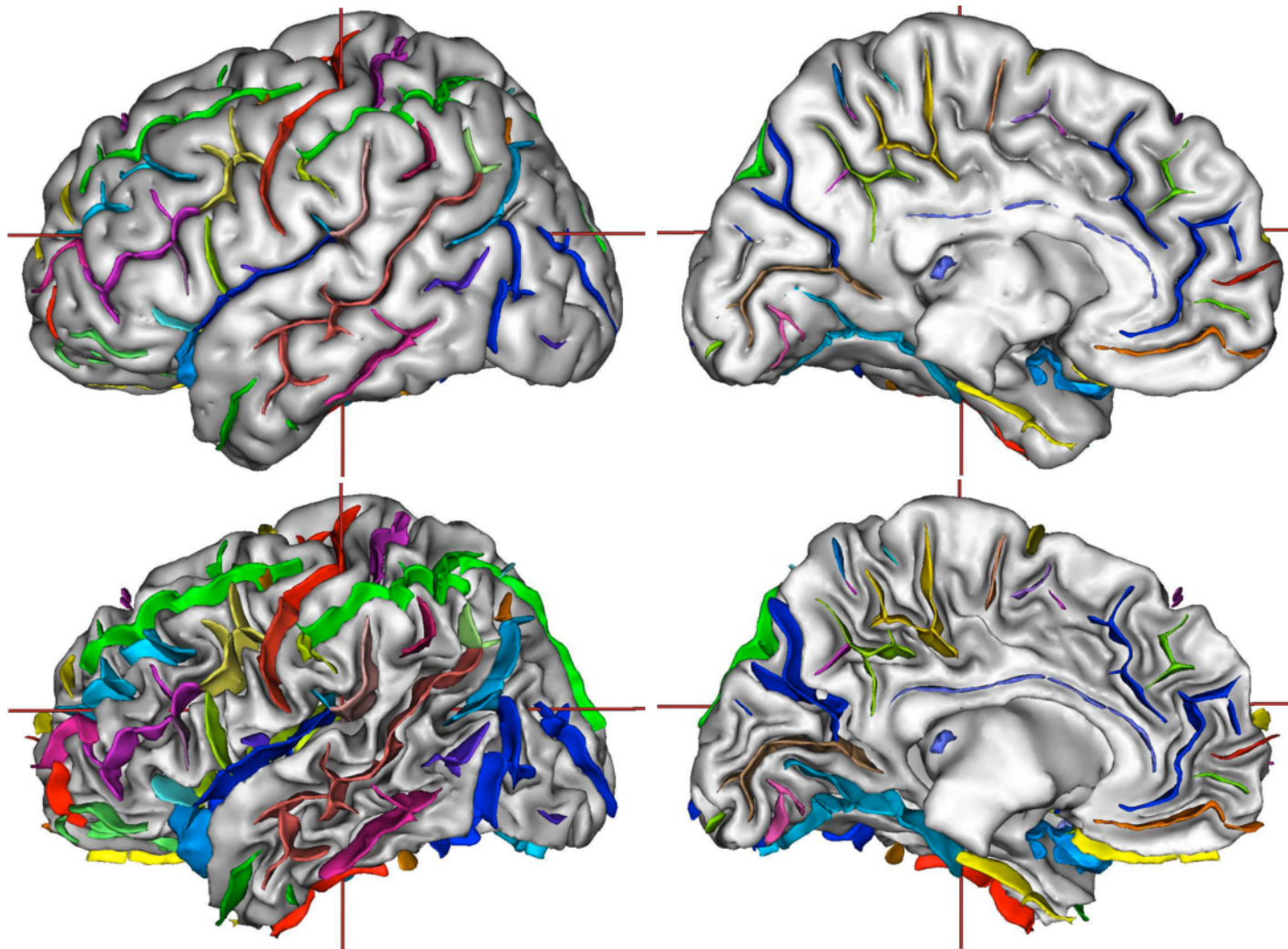
Candidate features in 3-D: sulcal basins



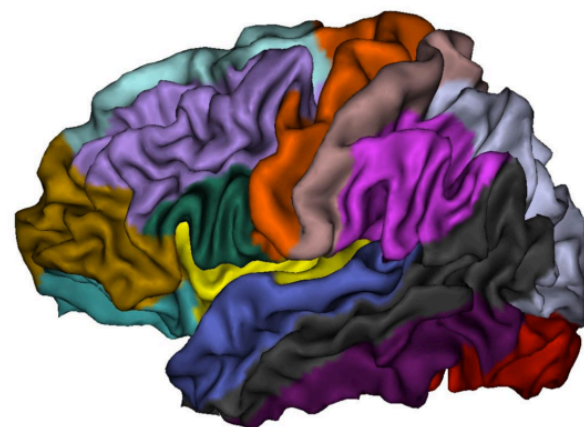
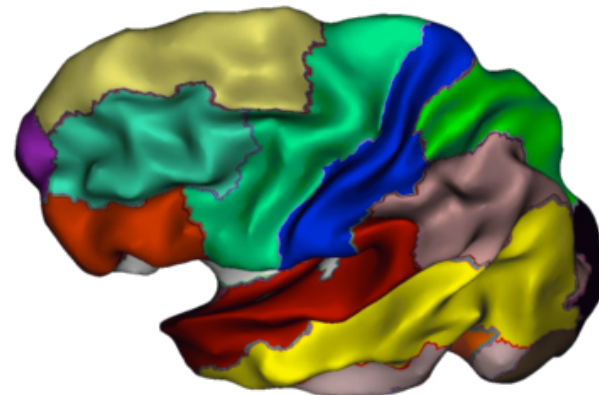
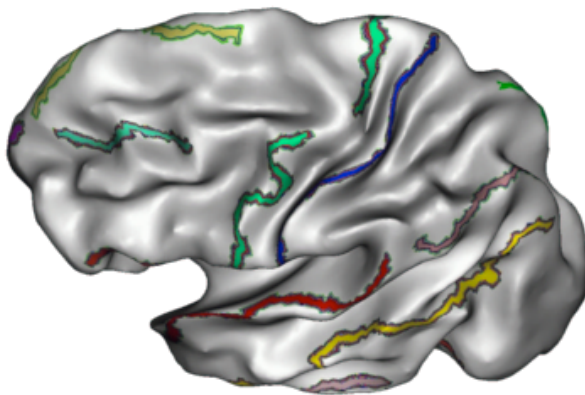
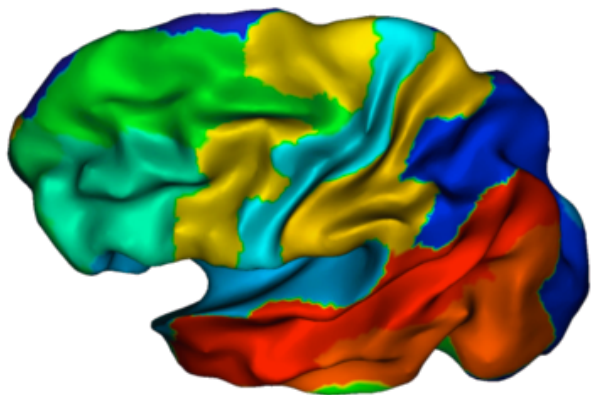
Candidate features in 3-D:
sulcal **skeletons**



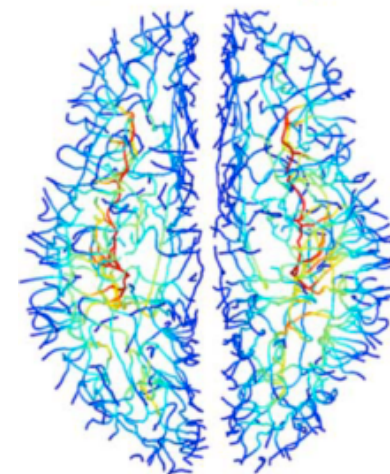
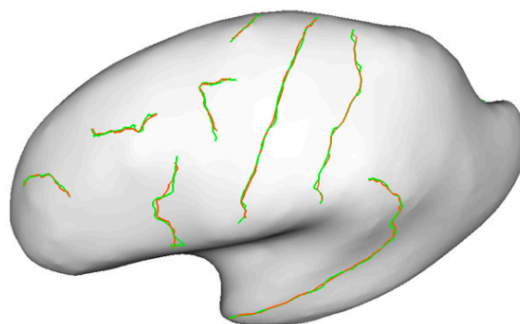
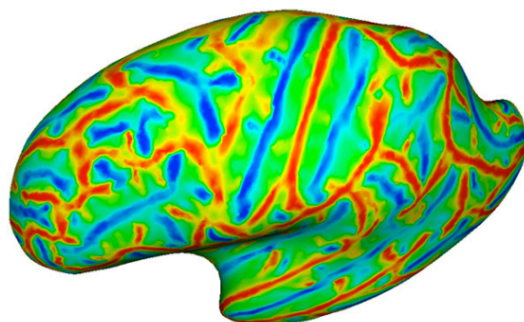
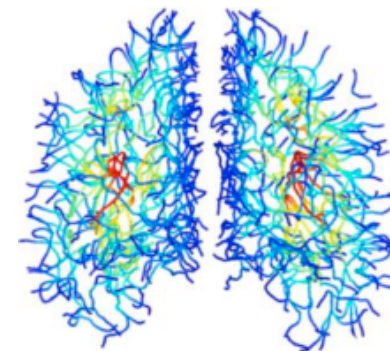
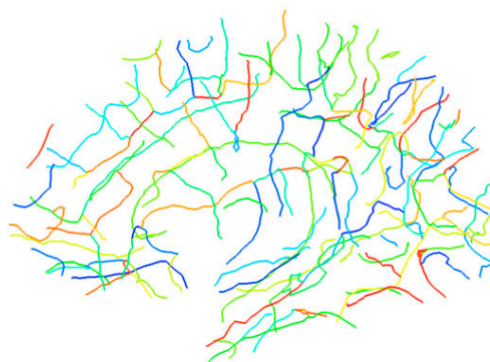
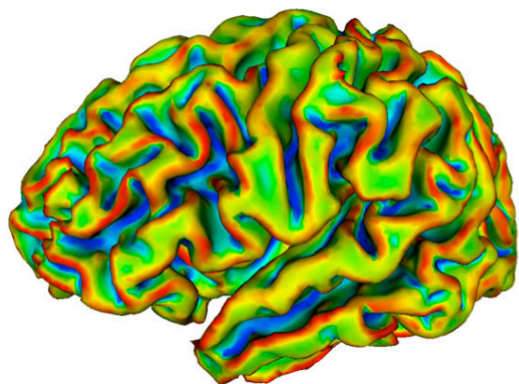
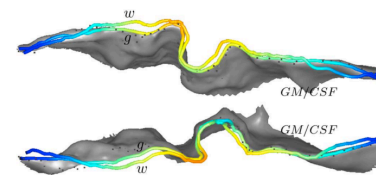
Candidate features in 2-D:
sulcal **ribbons**



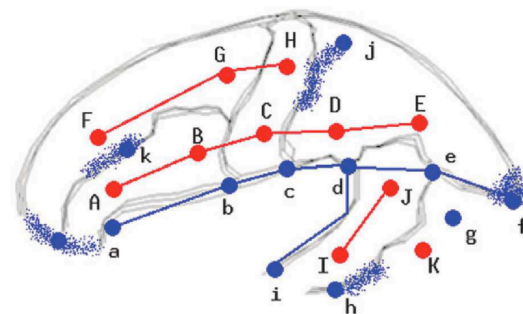
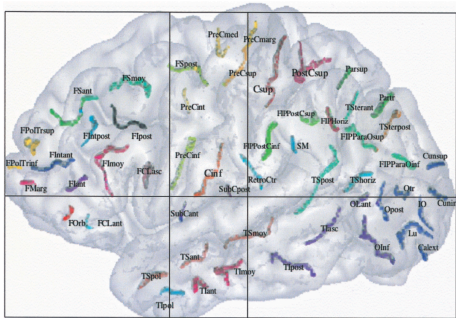
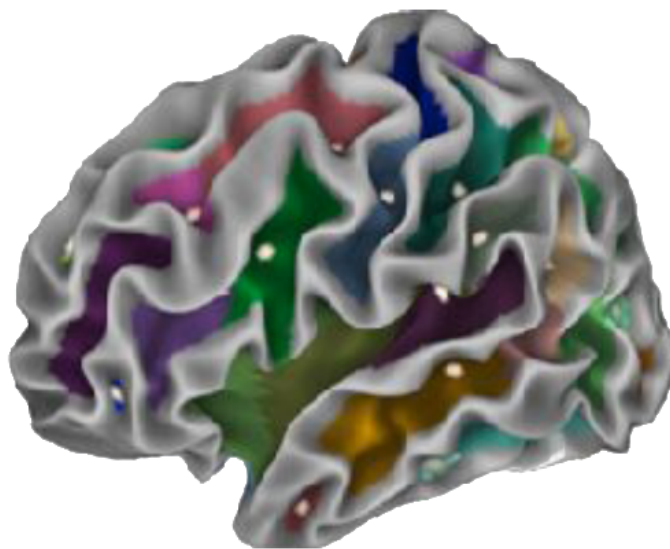
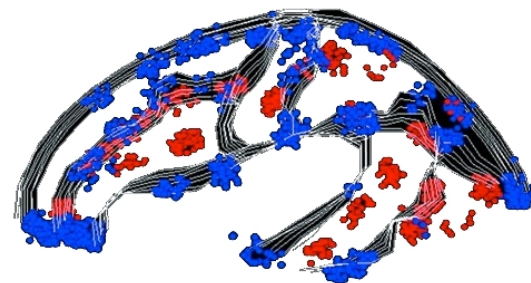
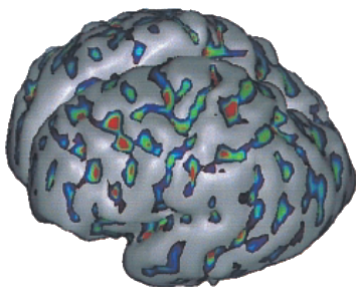
Candidate features in 2-D:
sulcal & gyral **surfaces**



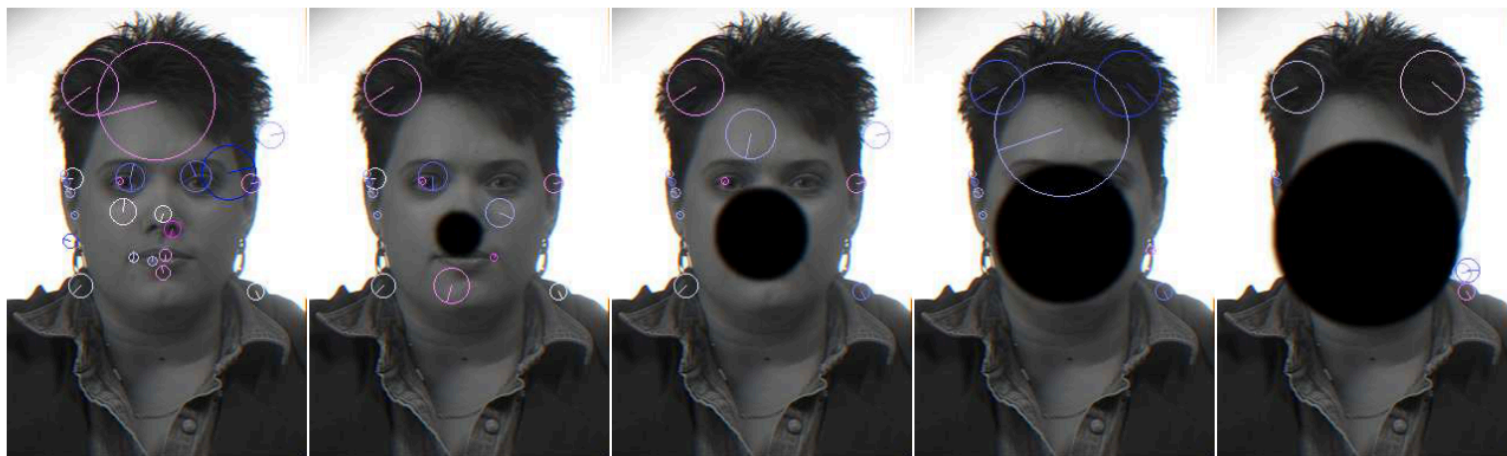
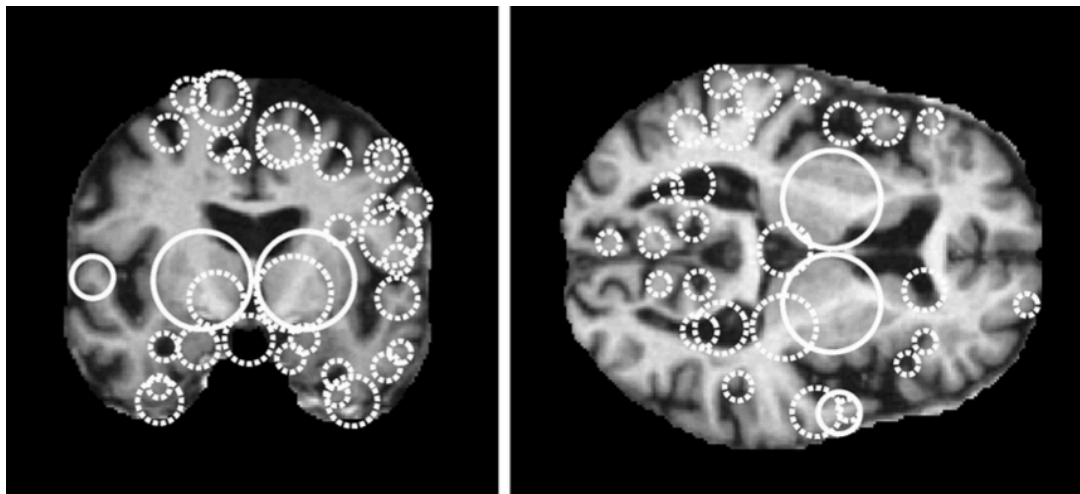
Candidate features in 1-D: sulcal **curves**



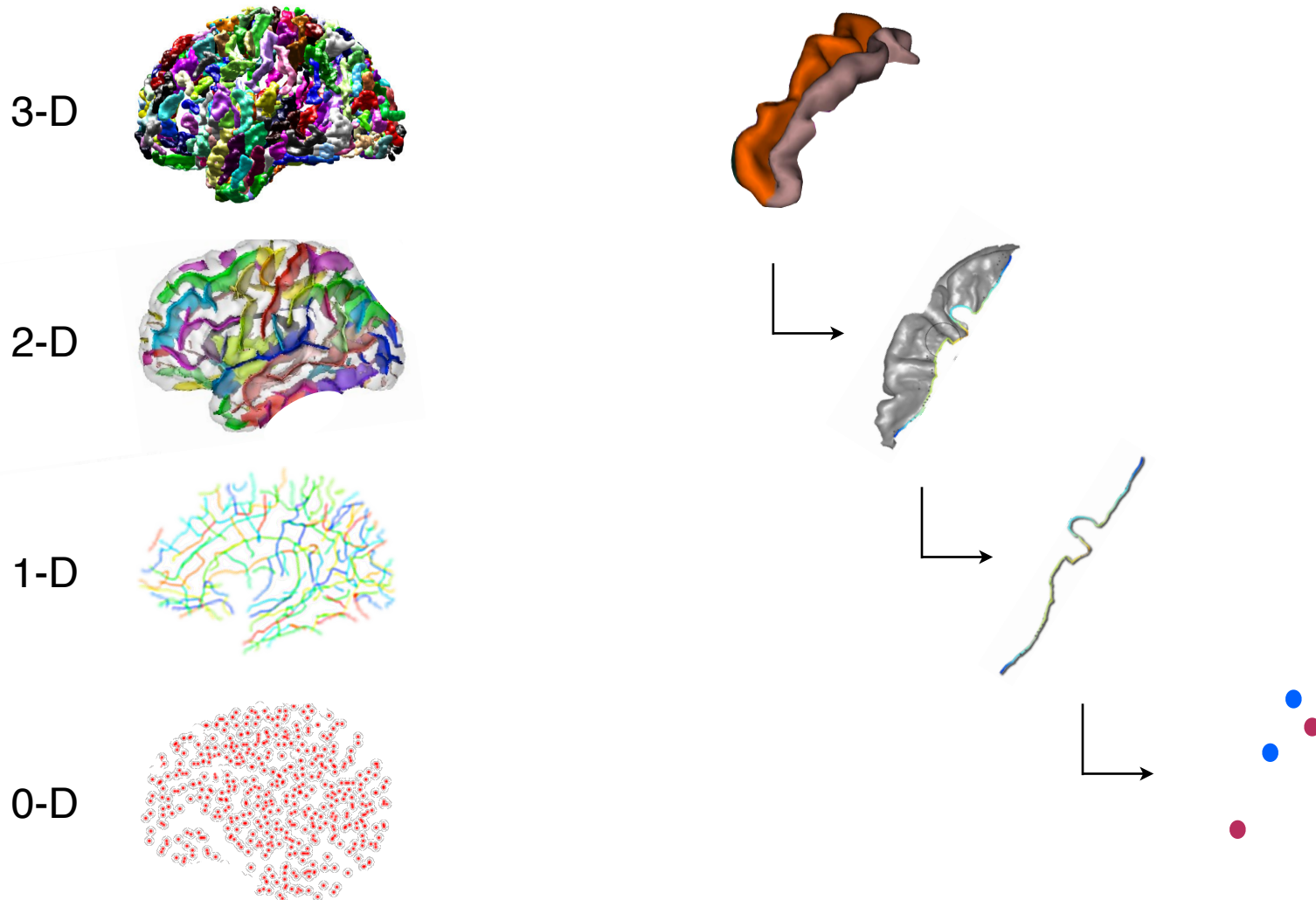
Candidate features in 0-D: sulcal pits



Candidate features in 0-D: SIFT points

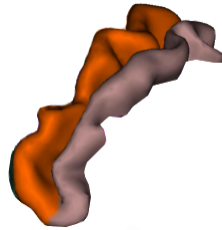
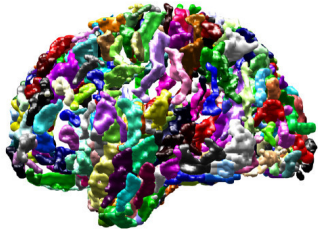


Proposed nested feature hierarchy



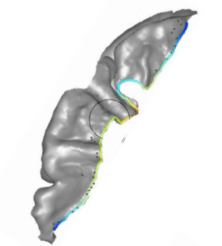
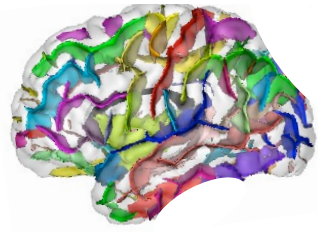
Candidate shape measures

3-D



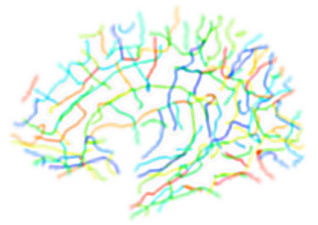
volume
surface area
lengths (thickness)
?

2-D



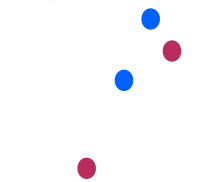
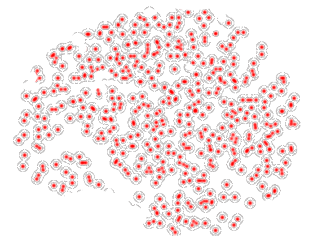
area
curvature
convexity
?

1-D



length
curvature
convexity
?

0-D

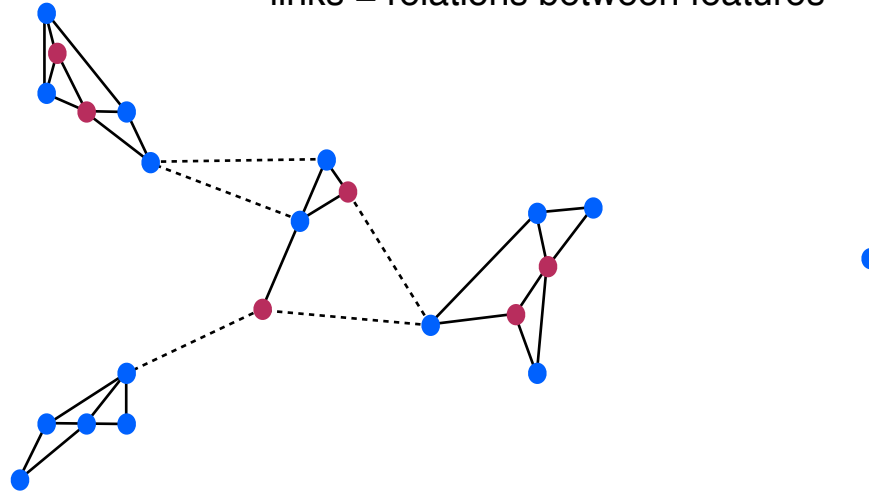


number of points
3-D convex hull volume
1-D sequence
?

Network analysis measures

nodes = features

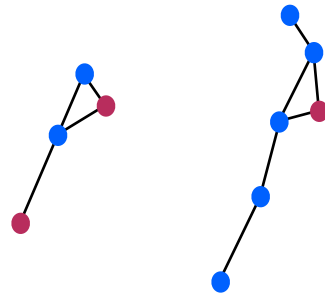
links = relations between features



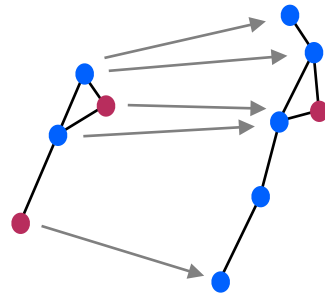
- centrality
- betweenness
- modularity

...

Deformation-based morphometry measures

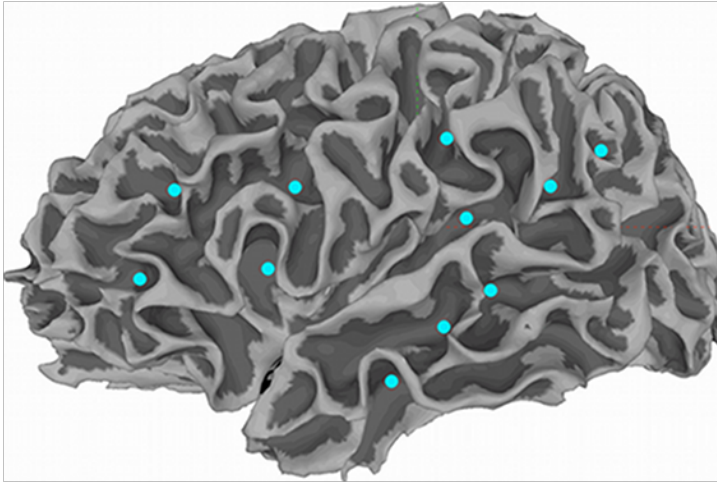


Deformation-based morphometry measures

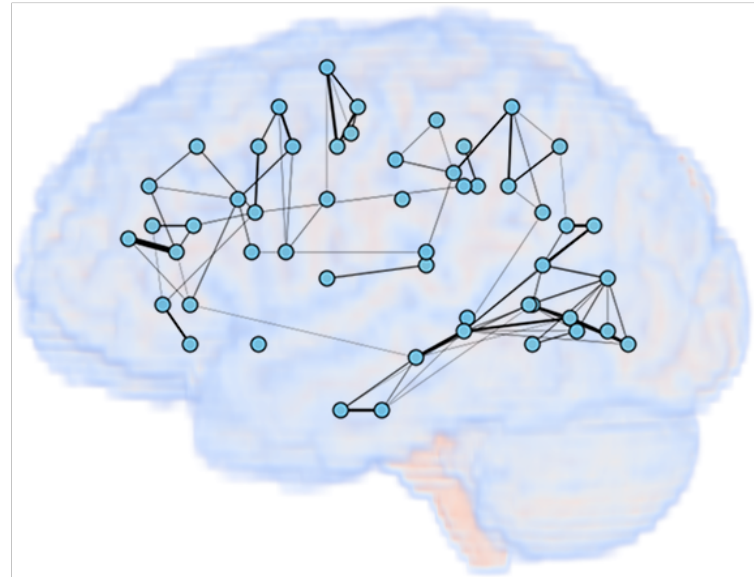


- warp field
- Jacobian
- Hausdorff distance

Multimodal feature graphs

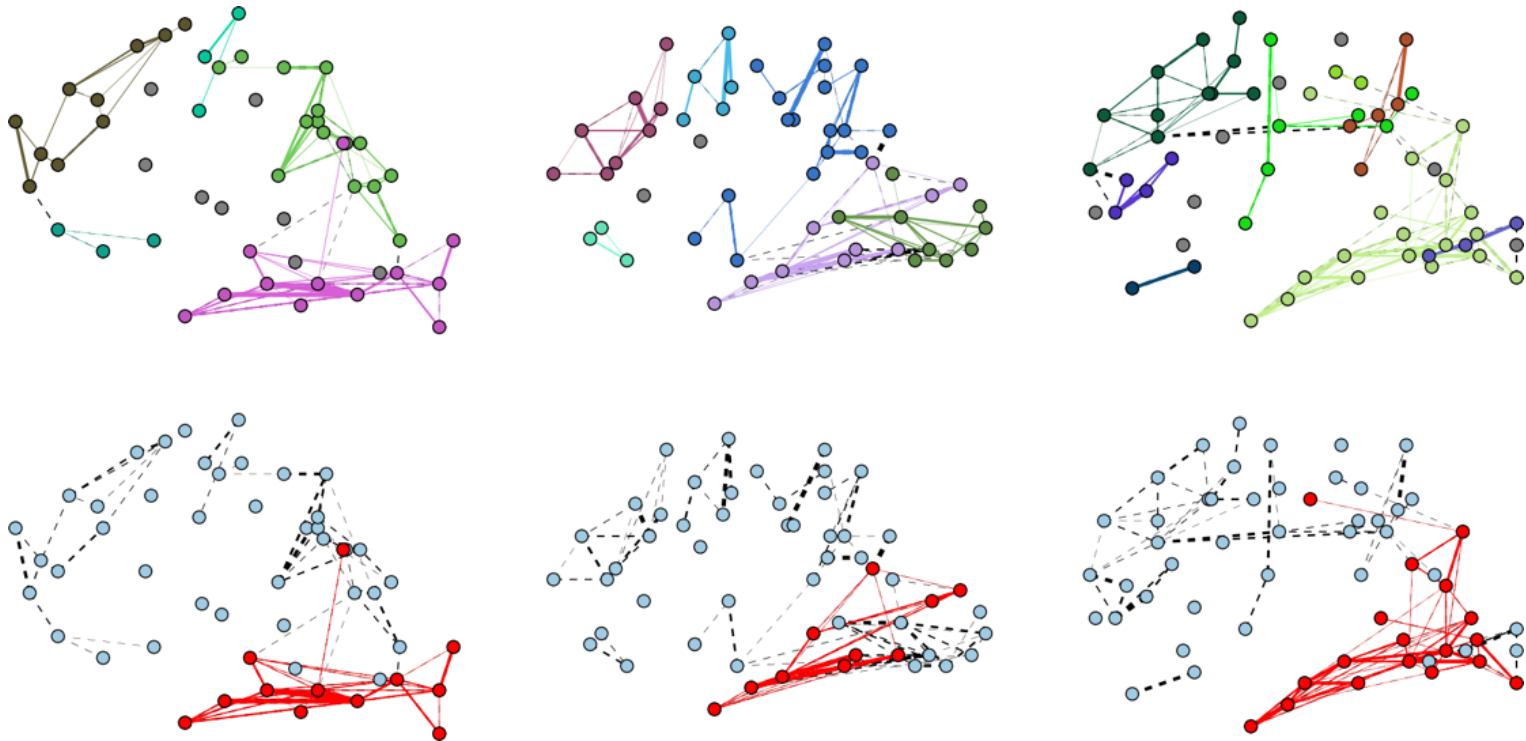


A gray/white matter surface (left lateral view) with visible sulcal pits highlighted (cyan circles). These features go by different names (sulcal roots, buried or annectant gyrii, plis de passage) and may be well conserved structures formed early in development.



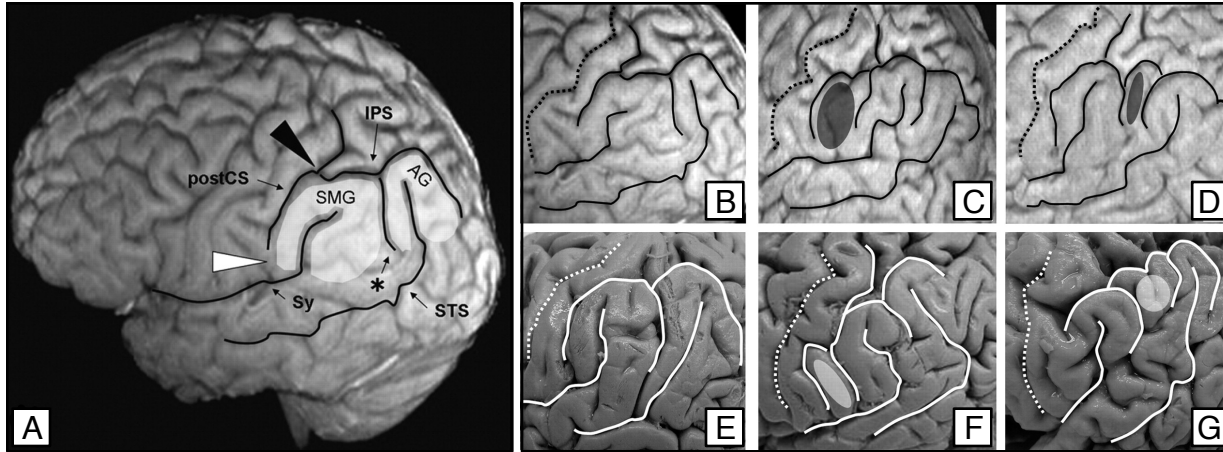
DTI connectivity graph (same subject). Vertices represent all extracted sulcal pits and each edge width indicates a connection probability greater than 0.01 between two vertices (and does not follow a tractography path).

Subgraph matching



Early attempt at subgraph extraction (upper row) and matching (lower row).
These graphs were constructed from (left to right) a remitter, non-remitter, and control subject.
The subgraphs in red in the lower row have the highest small-worldness ratio.

Anatomical variability



Example of natural morphological variability: left inferior parietal lobule (IPL; Kiriya et al. 2009).

(A-D) are MRI data and (E-G) are post-mortem specimens.

(A) IPL is highlighted and folds are outlined.

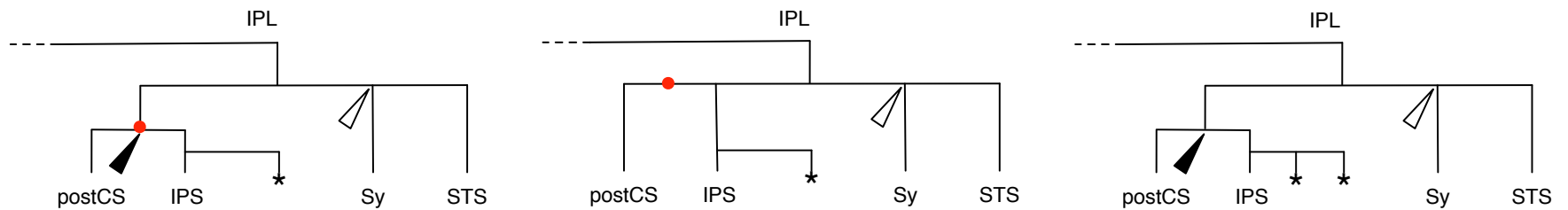
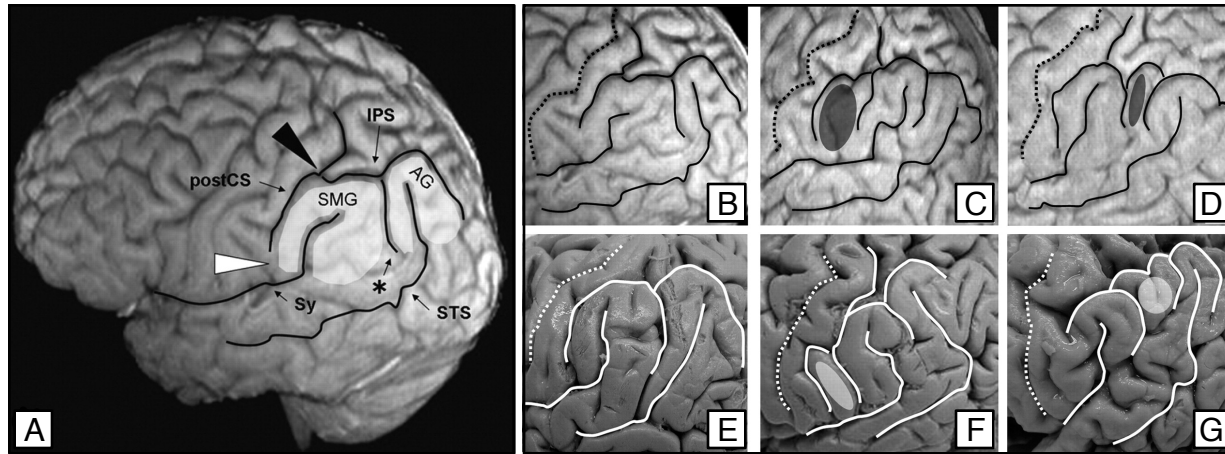
(B,E) Typical folding pattern.

(C,F) PreSMG pattern: an additional gyrus (ellipse) lies between postCS and SMG.

(D,G) PreAG pattern: an additional gyrus (ellipse) lies between SMG and AG.

[SMG: supramarginal gyrus; AG: angular gyrus; postCS: postcentral sulcus; IPS: intraparietal sulcus; Sy: Sylvian fissure, STS: superior temporal sulcus; *sulcus intermedius primus]

Graph-based representation of anatomical development



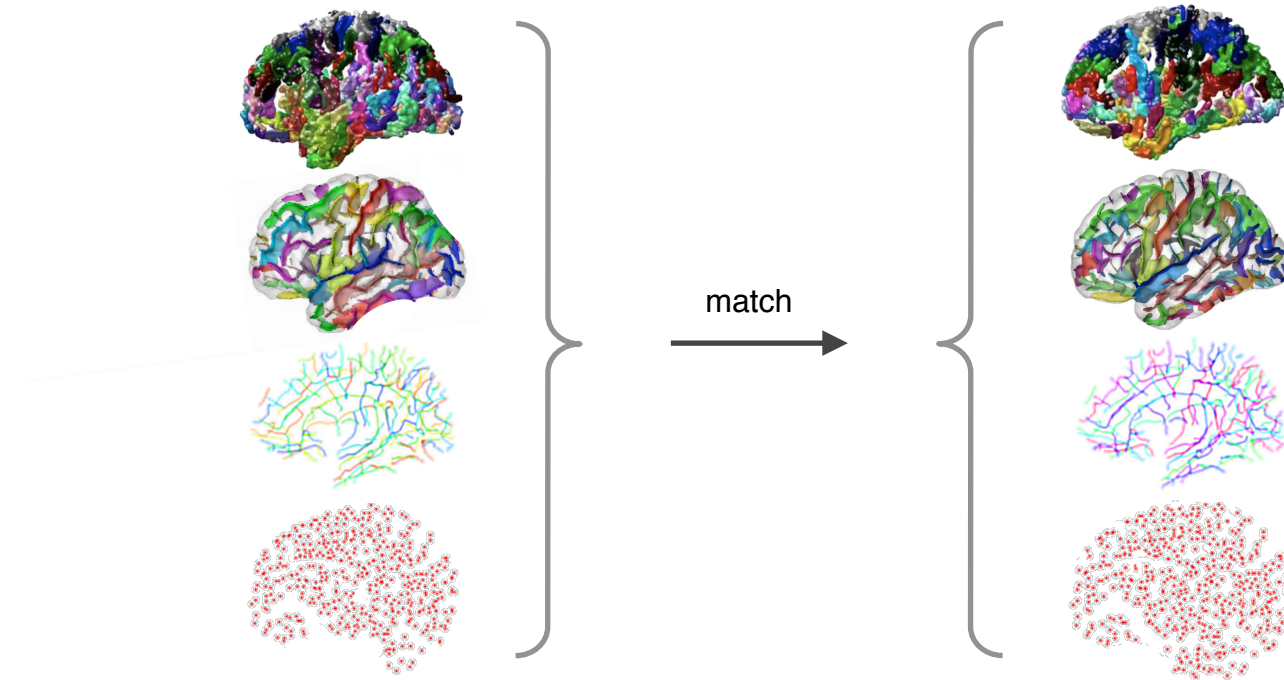
Feature matching

Bayesian face recognition algorithm

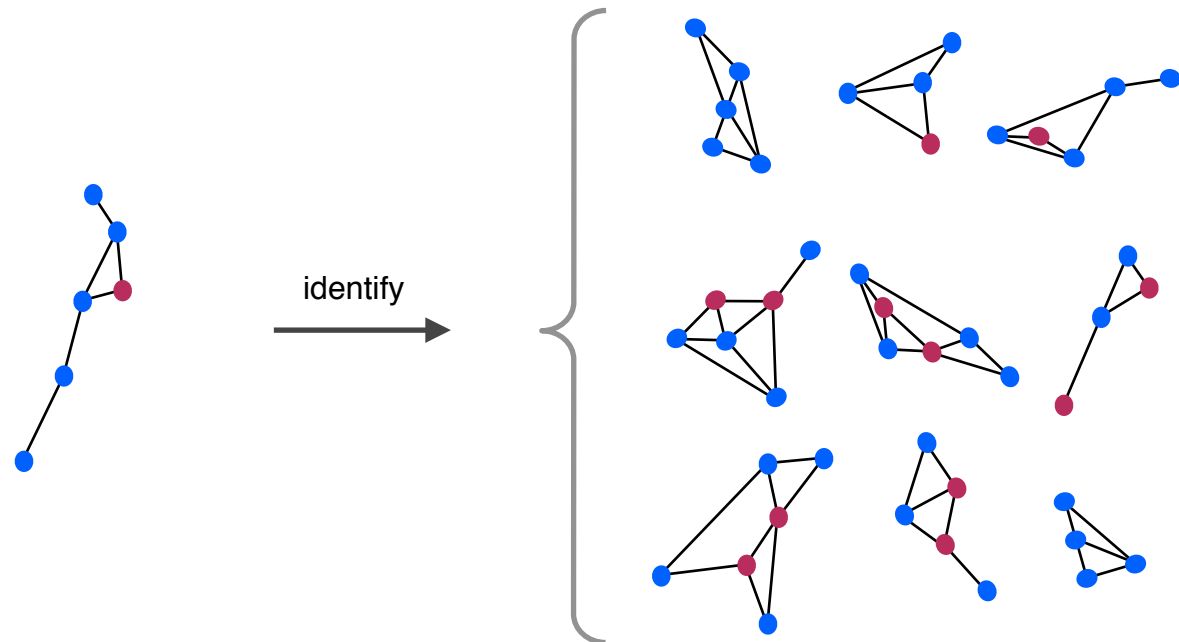
Example: MAP binary classification
(Moghaddam et al., 1998)

Feature matching

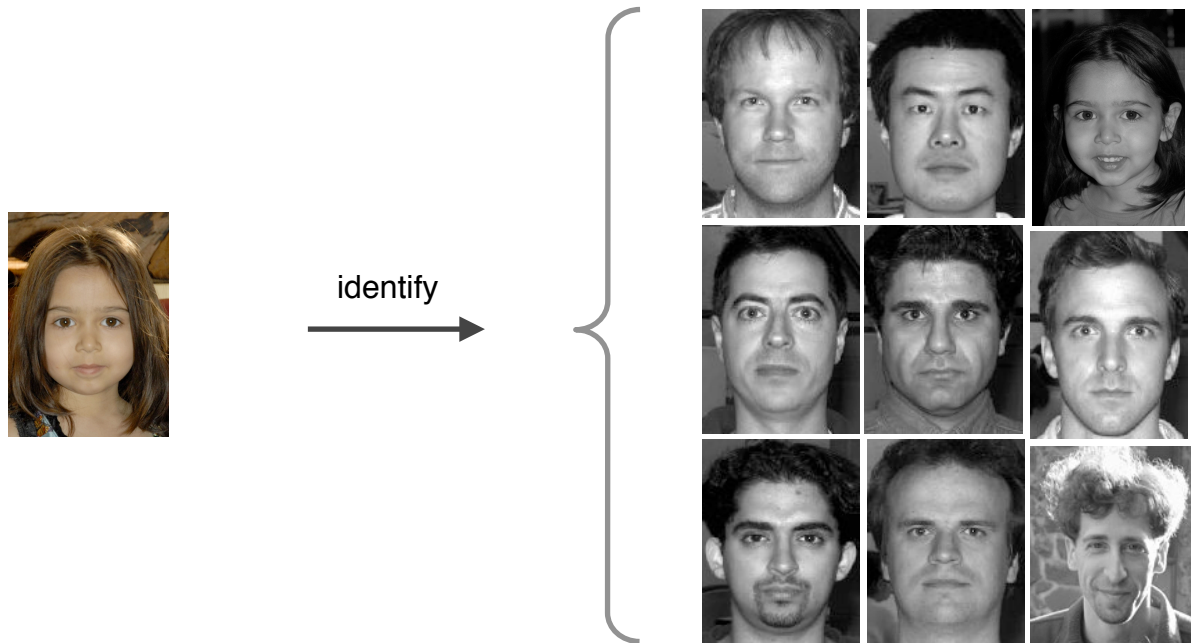
Instead of matching the entire set
of features across whole brains ...



... we can try to identify a group of features within a brain.



To adapt Bayesian face recognition to this problem, we can think about a set of facial features instead of a group of brain features ...

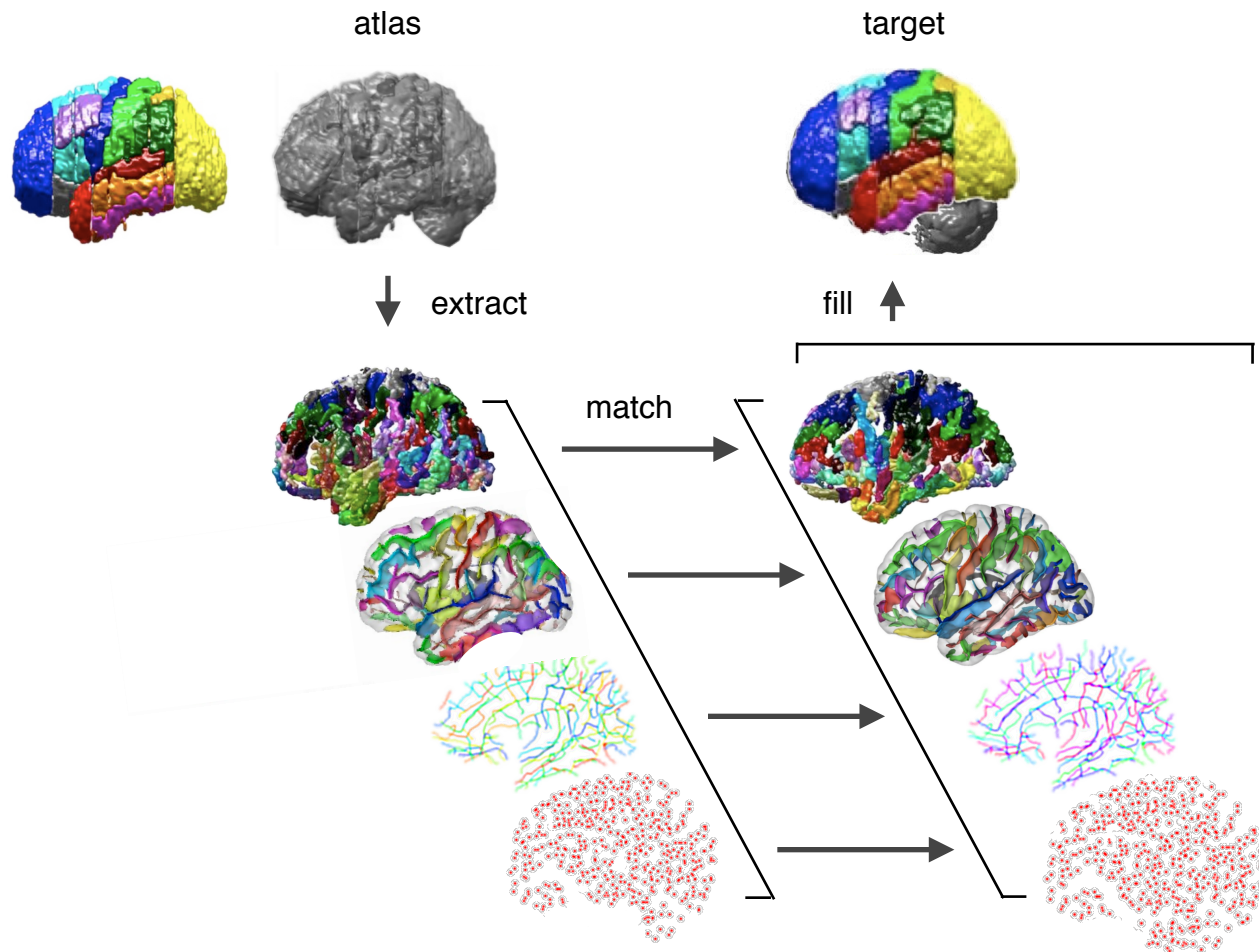


... and perform Bayesian binary classification,
by maximizing the posterior:



$$P(\Omega_I|\Delta) = \frac{P(\Delta|\Omega_I)P(\Omega_I)}{P(\Delta|\Omega_I)P(\Omega_I) + P(\Delta|\Omega_E)P(\Omega_E)}$$

Mindboggle 2: feature extraction feature matching feature-driven labeling





Denis Peruzzo
DTI tractography



Forrest Bao
(Texas Tech)
Nested feature extraction



Noah Lee
*Convolutional neural networks,
distributed database architecture*



Ray Razlighi
SIFT algorithm evaluation



Satrajit Ghosh (MIT)
*Machine learning,
NiPype software pipeline*



Steve Ellis
Computational topology